



DIU Génétique et Reproduction

Exercices NGS Séquençage ciblé

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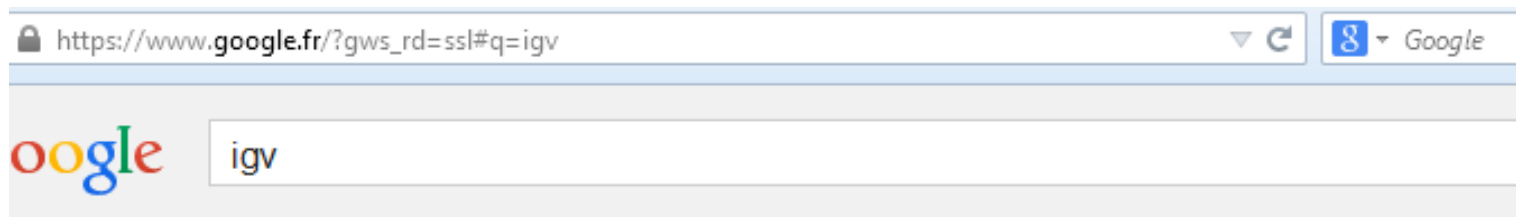
abdelkader.heddar@aphp.fr

Exercices

Analyse d'exomes

Récupérer sur <http://pdessen.free.fr/KB>

IGV Integrative Genomic Viewer



Web Images Actualités Vidéos Maps Plus ▾ Outils de recherche

Environ 6 870 000 résultats (0,47 secondes)

[Home | Integrative Genomics Viewer - Broad Institute](#)

www.broadinstitute.org/igv/ ▾ Traduire cette page

The IGV iPad app can now be installed from the Apple App Store. IGV for iPad is a lightweight genomic data viewer that provides some of the functionality ...

Downloads

If you have already registered for IGV please enter your ...

Register

IGV Registration. IGV is an open-source application, released ...

IGV User Guide

User Interface - Starting IGV - Viewing Alignments - ...

Viewing Alignments

The preferred file format for viewing alignments in IGV is the ...

Starting IGV

Starting IGV. You can start IGV from either the: Java Web Start ...

IGVTools

igvtools. The igvtools utility provides a set of tools for pre ...

[Autres résultats sur broadinstitute.org »](#)

<http://software.broadinstitute.org/software/igv/>

Install IGV :

<http://software.broadinstitute.org/software/igv/download>

Windows

1. Download the Windows package and execute the self-extracting archive.
2. It will prompt you for a location to extract the folder, choose anywhere you like (e.g. your home folder).
3. On completion, open the new folder.
4. Double-click the file "igv.bat", it might appear as just "igv" depending on your settings.

Download
Windows Package

Unzip dans un répertoire E:Archives_windows
IGV_2.3.82_jre.exe

Exécuter en cliquant sur IGV.bat

On peut aussi (voir page suivante) charger la Java Web Start

Par ailleurs

Récupérer une archive contenant les fichiers bam sur :

<http://pdessen.free.fr/KB/BAM.zip>

Désarchiver ce fichier (clic droit et extraction) sur le bureau

Java Web Start (All Platforms)

The buttons below use Java Web Start (JWS) to install and launch IGV directly from our web site.

***Mac Users:** The Java Web Start option does not work for some users due to security settings. The recommended solution is to use the bundled Mac App from the link above. Alternatively you can try to work around this by right-clicking on the buttons and saving the "jnlp" file, then right-clicking on the saved "jnlp" file and select "Open With > Java Web Start".

Chrome: Chrome does not automatically launch the Java Web Start files by default. Instead, the launch buttons below will download a "jnlp" file. This should appear in the lower left corner of the browser. Double-click the downloaded file to run, or if on a Mac right-click and select "Open With > Java Web Start"

Windows users: To run with more than 1.2 GB of memory you must install 64-bit Java. ***Most Windows installs do not include 64-bit Java by default, even if the operating system is 64-bit.*** Attempting to use the 2GB or greater launch options with 32-bit Java will result in the error *"could not create virtual machine"*.

 Launch with 750 MB	 Launch with 1.2 GB Maximum usable memory for Windows OS with 32-bit Java.	 Launch with 2 GB Maximum usable memory for 32-bit MacOS.	 Launch with 10 GB For large memory machines with 64-bit Java.
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BAM.zip (sur <http://pdessen.free.fr/KB/BAM>)

Unzip BAM.zip

BAM

test_HCV:

B00G39Y_chr16.bam

B00G39Y_chr16.bam.bai

B00G39Y_chr5.bam

B00G39Y_chr5.bam.bai

test_Tumor_Ctrl:

control

control_sorted.bam

control_sorted.bam.bai

QC_filtered

QC_raw

tumor

QC_filtered

QC_raw

tumor_sorted.bam

tumor_sorted.bam.bai

Mutations dans des gènes impliqués dans la reconnaissance du virus HCV (étude le M. MISRAHI)

Pour des questions de volume on a limité les données d'exome
à 2 bandes chromosomiques 5q13.2 et 16p13.3

Rappel : Seuls les zones couvrant les CDS et les bornes sont
Couvertes par des reads.

Un clic droit de souris fait apparaitre un grand nombre d'options

Dans IGV : **Sélectionner le build hg19**
 Charger le fichier B00G39Y_chr5.bam

Sachant qu'il y a des reads d'un exome autour de la bande 5q13.2
balayer cette zone pour trouver des gènes séquencés en suivant la piste
bam coverage

Quels gène(s) ? (autour de 68.800 Mb)
Combien d'exons ? (passer la souris sur le gène)
Sens du gène ?
Combien de transcrits ? (option squished sur refSeq)
Zoomer pour regarder les reads ?

Est-ce que tous les exons sont couverts ? CDS ?
Profondeur ?
Qualité ?
Trouver la (les) mutations et localiser les à la base près
Proportion d'allèles REF et ALT
Repérer la couleur différente sur le coverage . Visualiser les fréquences
Variant homozygote ou hétérozygote ?

Examiner les possibilités avec la touche Ctrl sur une position
Comment extraire une séquence locale ?

Dans IGV : Charger le fichier B00G39Y_chr16.bam

Sachant qu'il y a des reads d'un exome autour de la bande 16p13.3
Balayer cette zone pour trouver des gènes séquencés en suivant la piste
bam coverage

Quels gène(s) ?
Combien d'exons ?
Sens du gène ?

Zoomer pour regarder les reads

Est-ce que tous les exons sont couverts ? CDS ?
Profondeur ?
Qualité ?
Trouver la (les) mutations et localisez les à la base pres
Proportion d'allèle REF et ALT
Repérer la couleur différente sur le coverage . Visualiser les fréquences
Variant homozygote ou hétérozygote ?

Ctrl sur une position ?
Comment extraire une séquence locale
Une autre mutation ?

Regarder l'impact de ces mutations

Dans dbSNP <https://www.ncbi.nlm.nih.gov/SNP/>

1000 Genomes <http://www.internationalgenome.org/>

EVS <http://www.internationalgenome.org/>

ExAC <http://exac.broadinstitute.org/>

gnomAD <http://gnomad.broadinstitute.org/>

Fréquence et présence dans les populations (1000G)

Nature de l'impact ?

Tester PolyPhen2 <http://genetics.bwh.harvard.edu/pph2/>

Tester Sift http://sift.jcvi.org/www/SIFT_chr_coords_submit.htm

Voir les annotations dans UniProt <http://www.uniprot.org>

Sélection par CTRL de la séquence consensus

chr16:3065097-3065673

NN
NN
NN
NN
NNNNNNNNNNNNNNNNNNNNNAAATGAATTTTAAATATC
CTCAGTCAAAGAAAAAATAGCAGGAGGCAGAAA
CAAAAGGTACGAACCCATCCCAAAGCTGTTGGGC
ACTGCCACTTCTGGATGGCTCTAGCGCCAGCGGA
GCCCCATTCCCCTCCACGTCAGACGTAATTCTTG
GTAGGGTACTCAGAGGGCCCCYGAGAGATGGCA
GGGGCAGATGTTGAGTAGCGGGCCATGTAATGGC
TGGGGCCCTGGGACCCCCCGAGGGGCAAGTGC
AGCACAGCAACCCCCCACCCAGCAACAAAAGGCC
TGAGGCCGCCCAGCCCAAGTAGAGGGAGGCCCC
CAGCTCCCGCTTTTGGGCCTCAGCCACCAGGGGG
TTATAGAAGTCCCGGATGACGGCATGCGCCGTCCA
GCACACGGGGATTAGCGTCAGGACCCCTGAGATG
ACAAAGACAATCCCAGAGGTGAGCACCA

Clic droit sur le polymorphisme
chr5:68804985 C>G Pro

chr5:68788119-68853931

OCLN

chr5:68788119-68853931

id = NM_002538

Exon number: 3

Amino acid coding number: 24

chr5:68804968-68805646

chr5:68,804,987

<hr>Total count: 144

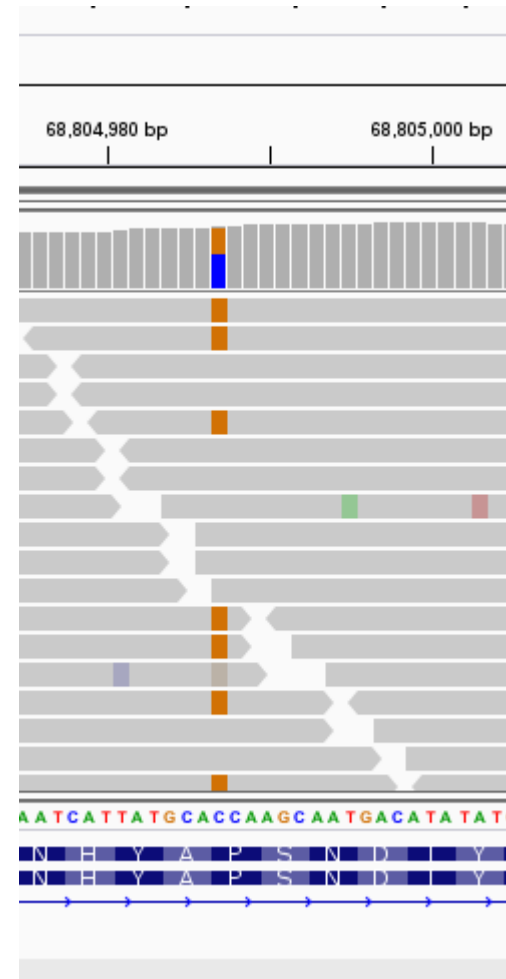
A : 0

C : 80 (56%, 58+, 22-)

G : 64 (44%, 40+, 24-)

T : 0

N : 0

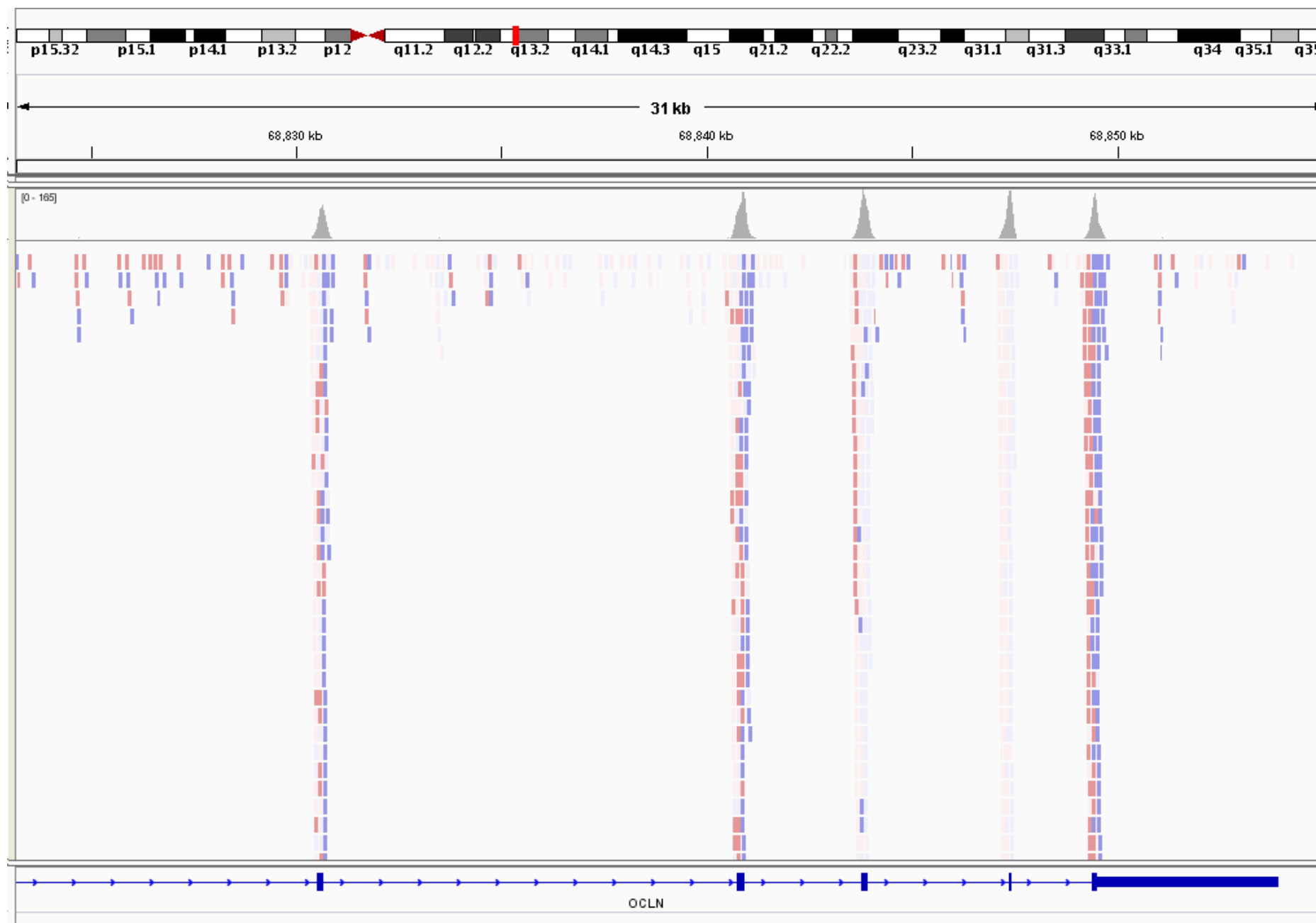


chr	beg	rc_snp	REF	ALT	EFFeffect	EFFImpact	EFFclass	EFFcodon	EFFaa	EFFsymb	B00G39X	B00G39Y
3	190030989	rs10513846	G	A	EXON	MODIFIER				CLDN1	1	0
16	3065397	rs149605777	C	T	NON_SYNONYMOUS_CODING	MODERATE	MISSENSE	cGg/cAg	R209Q	CLDN6	0	1
16	3065635	rs138377300	C	G	NON_SYNONYMOUS_CODING	MODERATE	MISSENSE	Gtc/Ctc	V130L	CLDN6	0	0
16	3065596	rs2257295	T	C	NON_SYNONYMOUS_CODING	MODERATE	MISSENSE	Atc/Gtc	I143V	CLDN6	1	2
16	3063149	rs2231608	T	G	START_GAINED	LOW				CLDN9	0	0
5	68804987	rs147125035	C	G	NON_SYNONYMOUS_CODING	MODERATE	MISSENSE	Cca/Gca	P24A	OCLN	0	1
5	68849586	.	G	A	EXON	MODIFIER				OCLN	0	0
5	68830632	.	C	T	NON_SYNONYMOUS_CODING	MODERATE	MISSENSE	Cgg/Tgg	R13W	OCLN	0	0

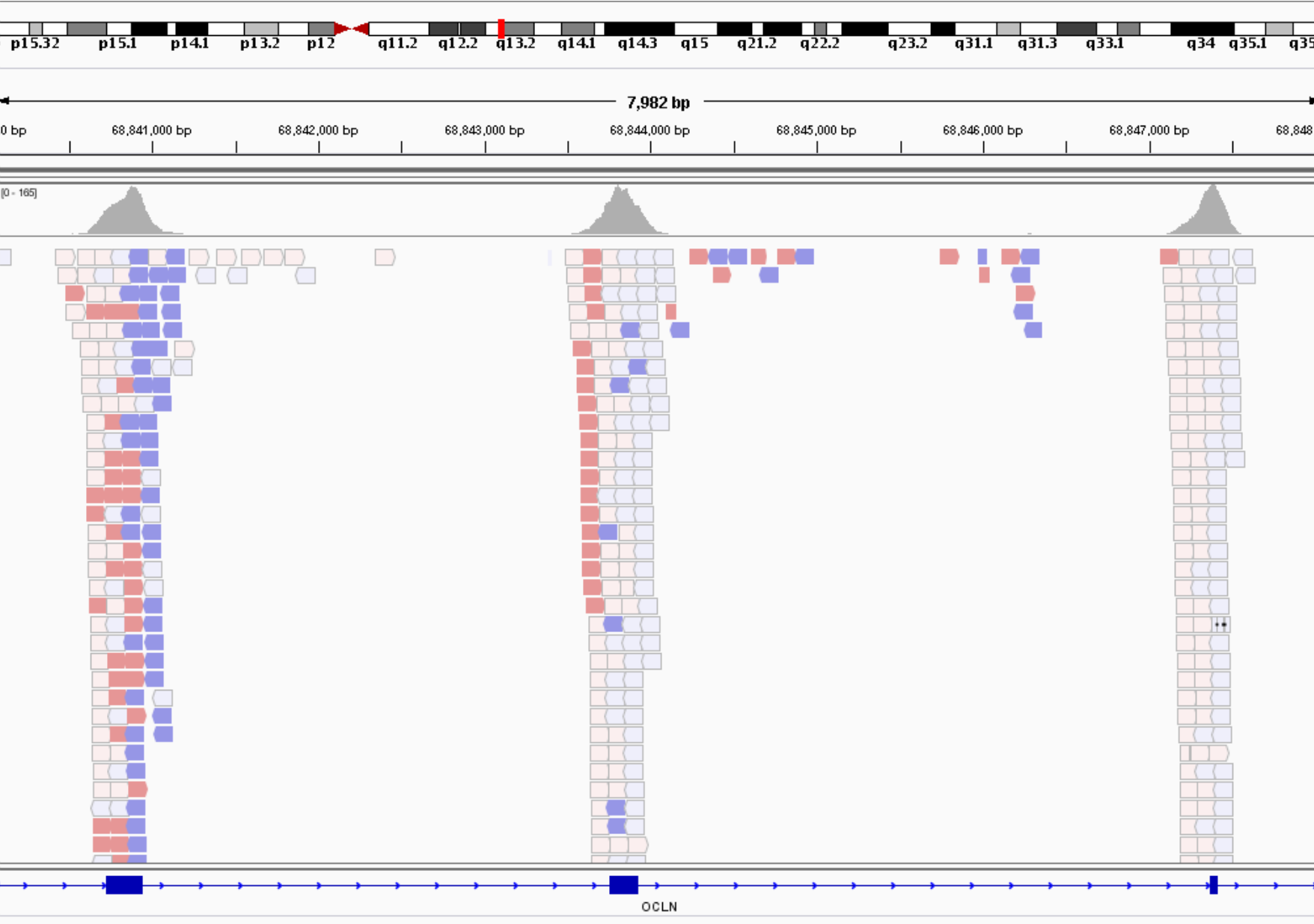
Visualisation des mutations (validation sur les gènes déjà séquencés en Sanger)

gène	mutation	dbSNP rs#ID		code
CLDN6	c.626 G>A (R209Q)	rs149605777	chr3:3065397	B00G39Y
OCLN	c.70 C>G (P24A)	rs147125035	chr5:68804987	B00G39Y
SR-B1	c.403 G>A	rs5891	chr12:125299542	B00G39Z
SR-B1	NC+23 C>T	NA	en 3'UTR ?	B00G39X
SR-B1	c.1056 G>A (V35I)	rs372663606	chr12:125284742	B00G39Z





B00G39Y OCLN



Couverture de capture Agilent v5

A partir du fichier bed : annotation des exons présents

```
$ more refg.txt
```

```
chr01:000011874-000012227__DDX11L1__NR_046018__exon__ex_01
chr01:000012613-000012721__DDX11L1__NR_046018__exon__ex_02
chr01:000013221-000014409__DDX11L1__NR_046018__exon__ex_03
chr01:000014362-000014829__WASH7P__NR_024540__exon__ex_11
chr01:000014970-000015038__WASH7P__NR_024540__exon__ex_10
```

820772 exons couverts

Dans refGene (hg19)

```
grep exon /db/GoldenPath/current/database/refGene.gff | wc -l
467234
```

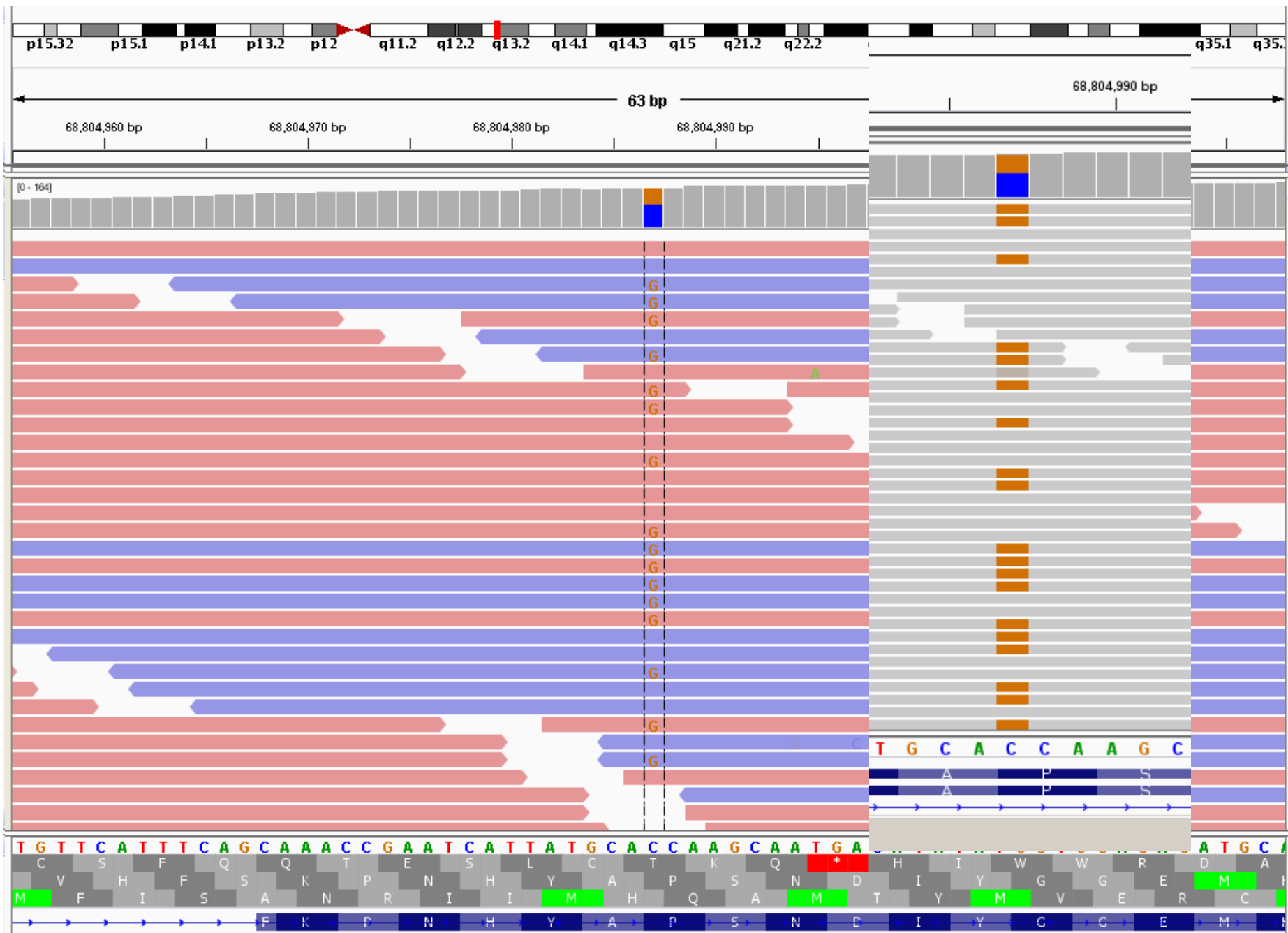
```
grep exon /db/GoldenPath/current/database/refGene.gff | awk '{ print $2}' | sort -u | wc -l
25981  nbre de genes
```

Genes absents

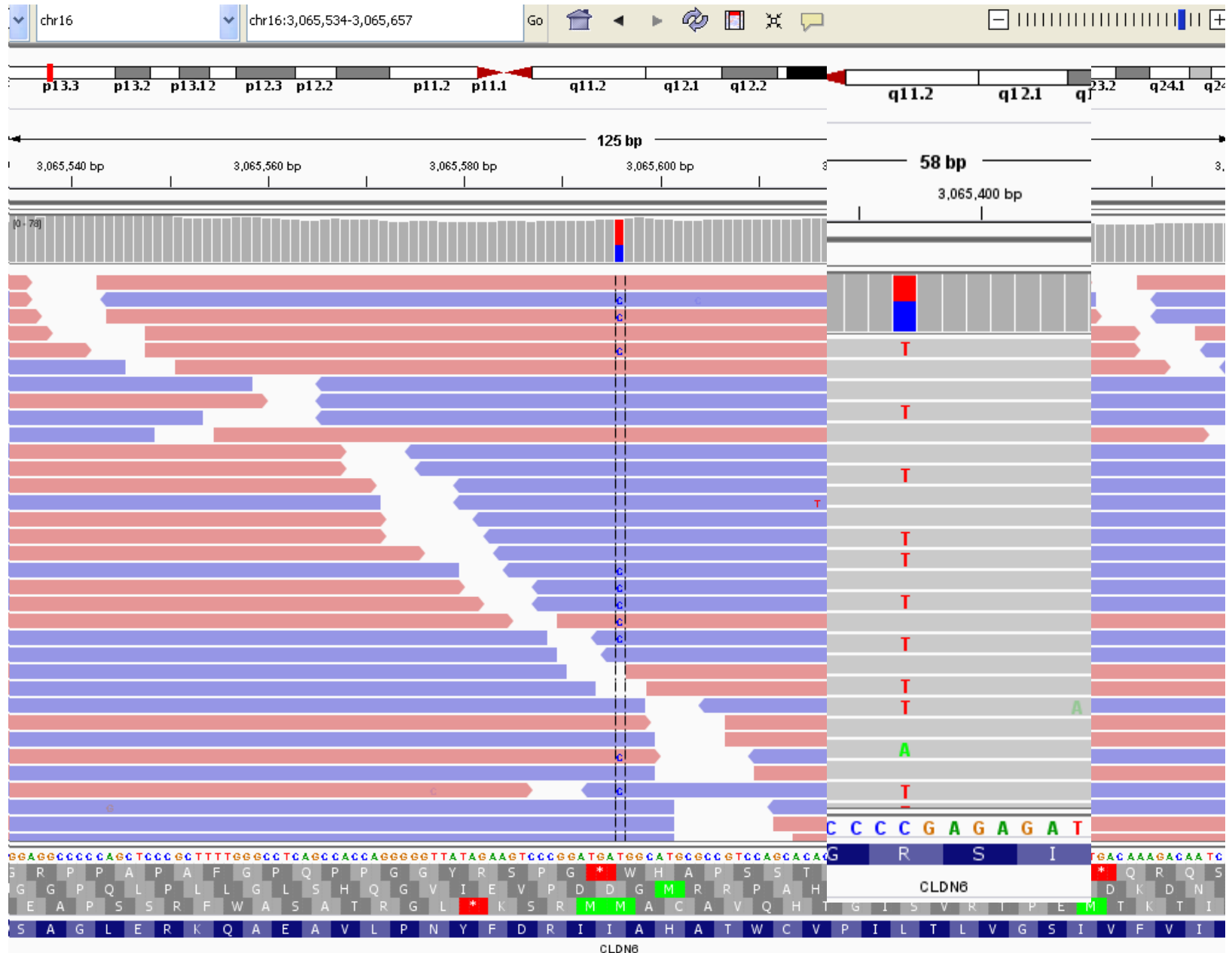
```
wc list_genes_CDS_absent.txt
```

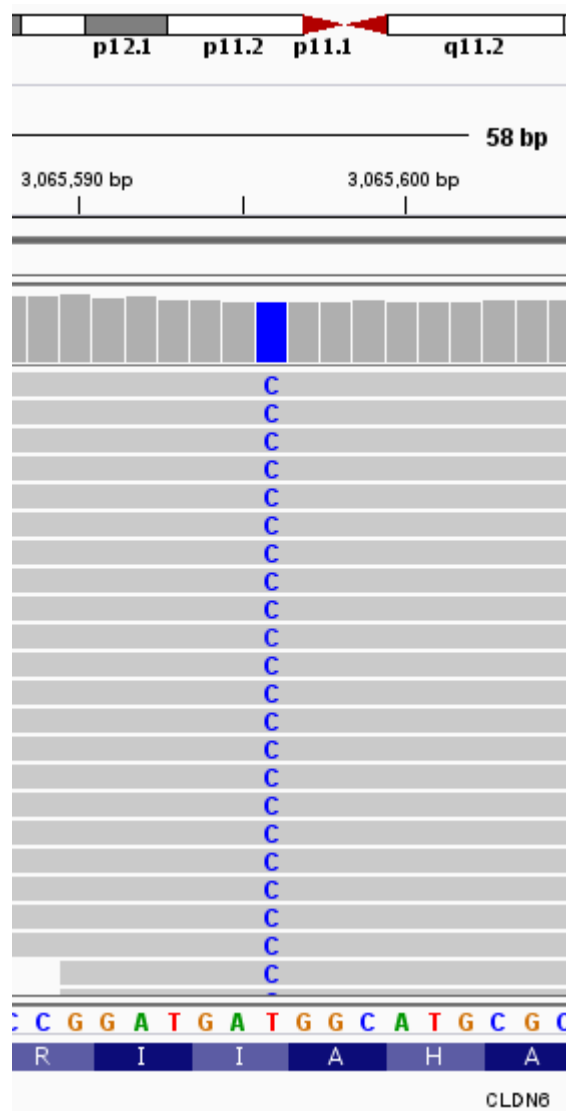
```
900 list_genes_CDS_absent.txt
```

(sur un seul sample)



CLDN6 chr16:3076596 T>C 28/42





Visualisation d'une mutation du gène PI3KCA sur un couple tumeur/normal

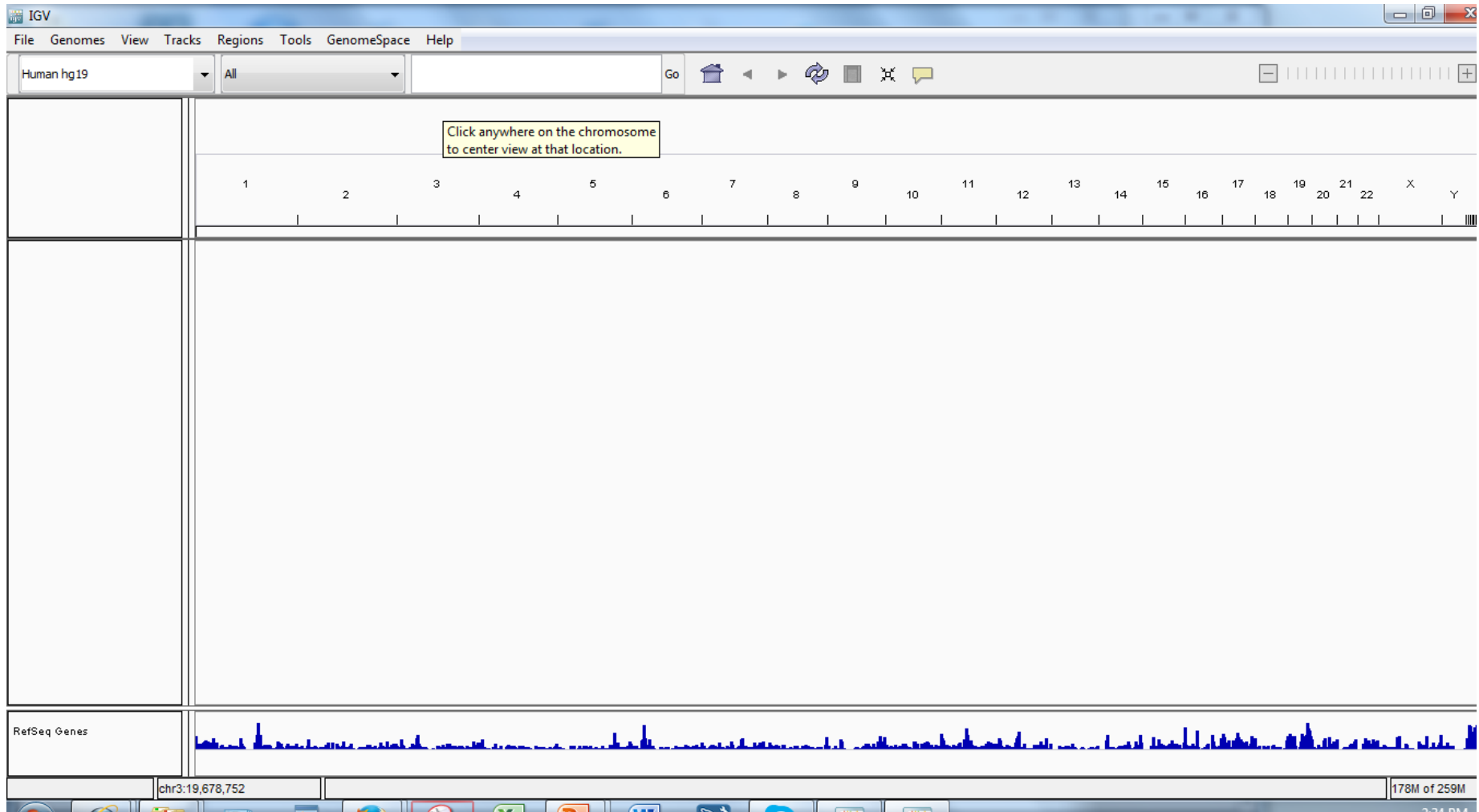
Sur IGV

Fichiers :
control.bam et tumor.bam

Sequencage ciblé autour du gène PIK3CA
(chr3:178Mb)

Menu genome : choisir human hg19

Puis les fichiers bam (indexés en bai)

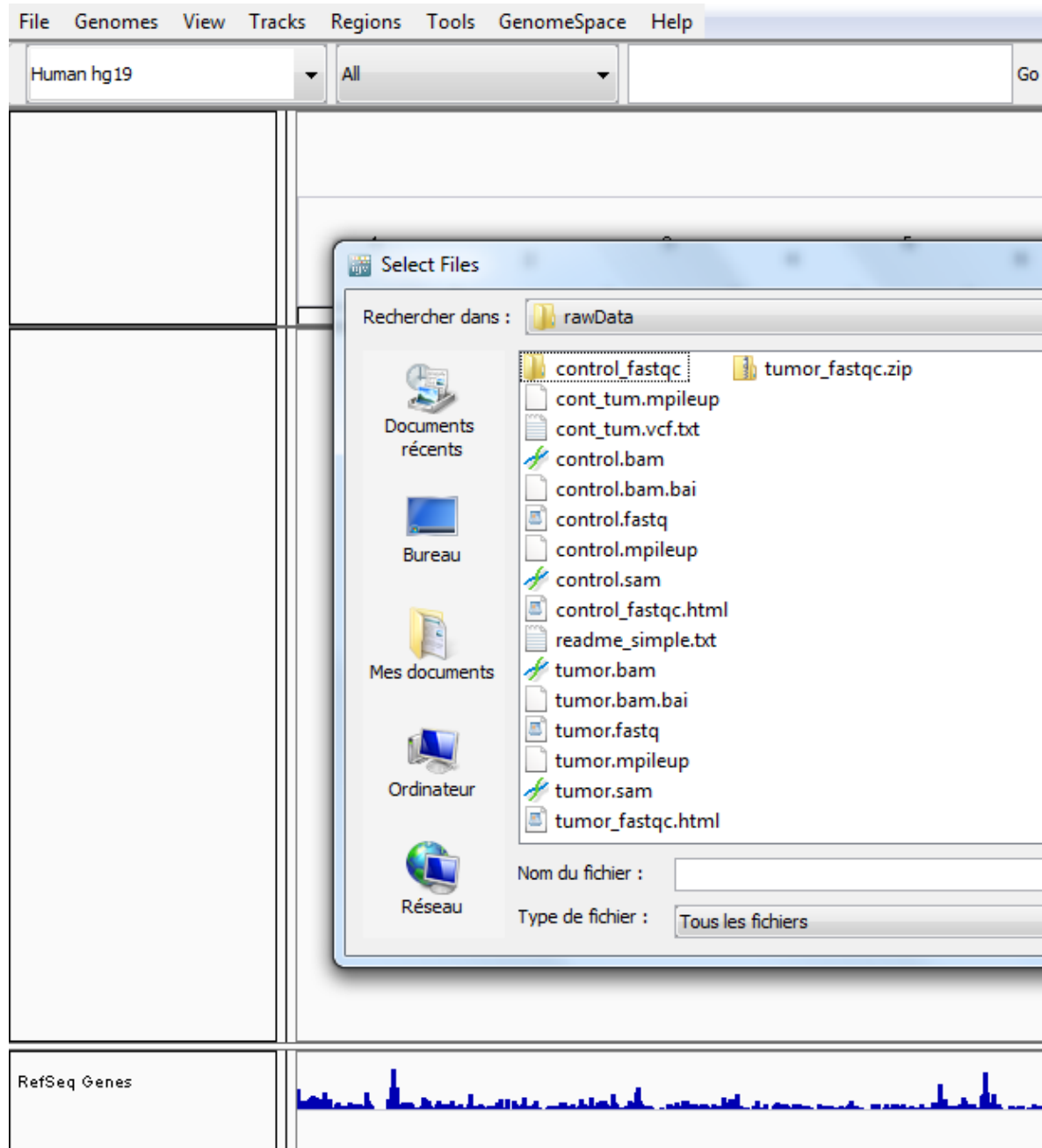


Piste de référence : RefSeq

sur hg19 : refGene.gff

NM_006218	PIK3CA	mRNA	chr3	178866311	178952497	.	+	.	21
NM_006218	PIK3CA	exon	chr3	178866311	178866391	ex_01	+	-1	.
NM_006218	PIK3CA	exon	chr3	178916538	178916965	ex_02	+	0	.
NM_006218	PIK3CA	exon	chr3	178917478	178917687	ex_03	+	1	.
NM_006218	PIK3CA	exon	chr3	178919078	178919328	ex_04	+	1	.
NM_006218	PIK3CA	exon	chr3	178921332	178921577	ex_05	+	0	.
NM_006218	PIK3CA	exon	chr3	178922291	178922376	ex_06	+	0	.
NM_006218	PIK3CA	exon	chr3	178927383	178927488	ex_07	+	2	.
NM_006218	PIK3CA	exon	chr3	178927974	178928126	ex_08	+	0	.
NM_006218	PIK3CA	exon	chr3	178928219	178928353	ex_09	+	0	.
NM_006218	PIK3CA	exon	chr3	178935998	178936122	ex_10	+	0	.
NM_006218	PIK3CA	exon	chr3	178936984	178937065	ex_11	+	2	.
NM_006218	PIK3CA	exon	chr3	178937359	178937523	ex_12	+	0	.
NM_006218	PIK3CA	exon	chr3	178937737	178937840	ex_13	+	0	.
NM_006218	PIK3CA	exon	chr3	178938774	178938945	ex_14	+	2	.
NM_006218	PIK3CA	exon	chr3	178941869	178941975	ex_15	+	0	.
NM_006218	PIK3CA	exon	chr3	178942488	178942609	ex_16	+	2	.
NM_006218	PIK3CA	exon	chr3	178943750	178943828	ex_17	+	1	.
NM_006218	PIK3CA	exon	chr3	178947060	178947230	ex_18	+	2	.
NM_006218	PIK3CA	exon	chr3	178947792	178947909	ex_19	+	2	.
NM_006218	PIK3CA	exon	chr3	178948013	178948164	ex_20	+	0	.
NM_006218	PIK3CA	exon	chr3	178951882	178952497	ex_21	+	2	.

Charger les fichiers .bam control et tumor

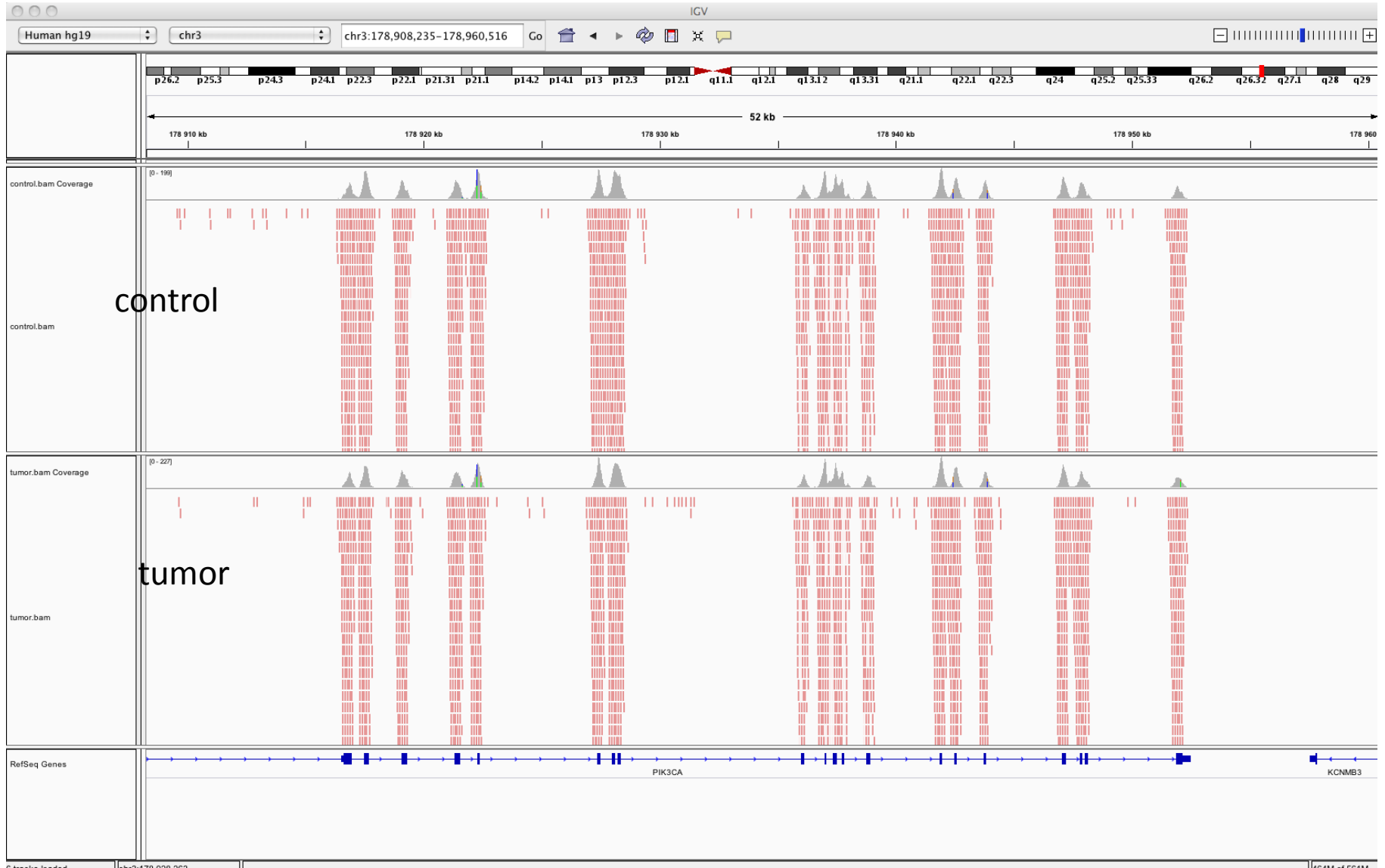


Regarder les menus
Utiliser le clic droit pour
Les options de visualisation

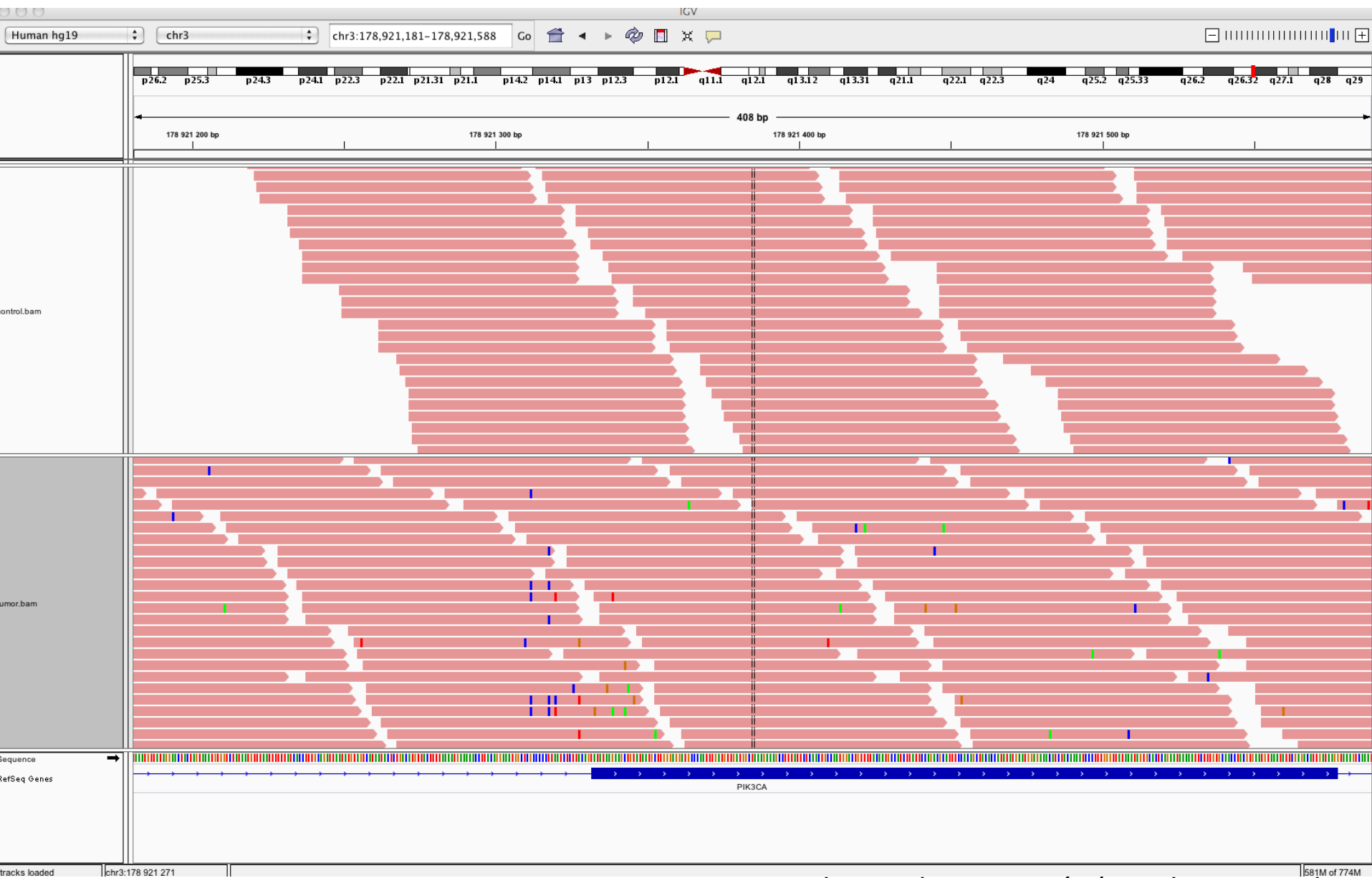
Déplacement des corrdonnées
Zoom

Les reads ne s affichent que si
Le zoom est suffisant

IGV 2.3 (hg19)

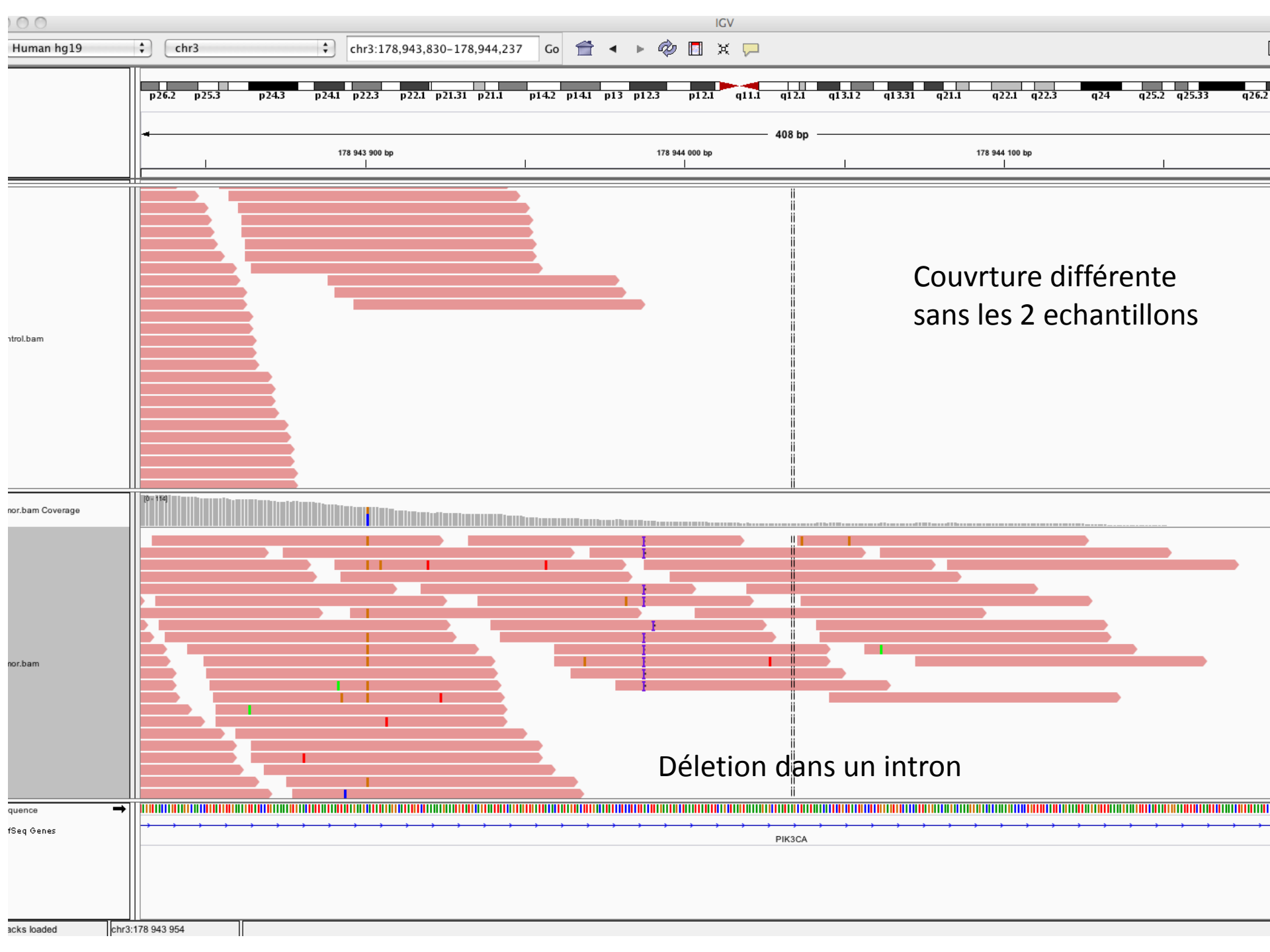


Exons du gène PIK3CA chr3:178900000

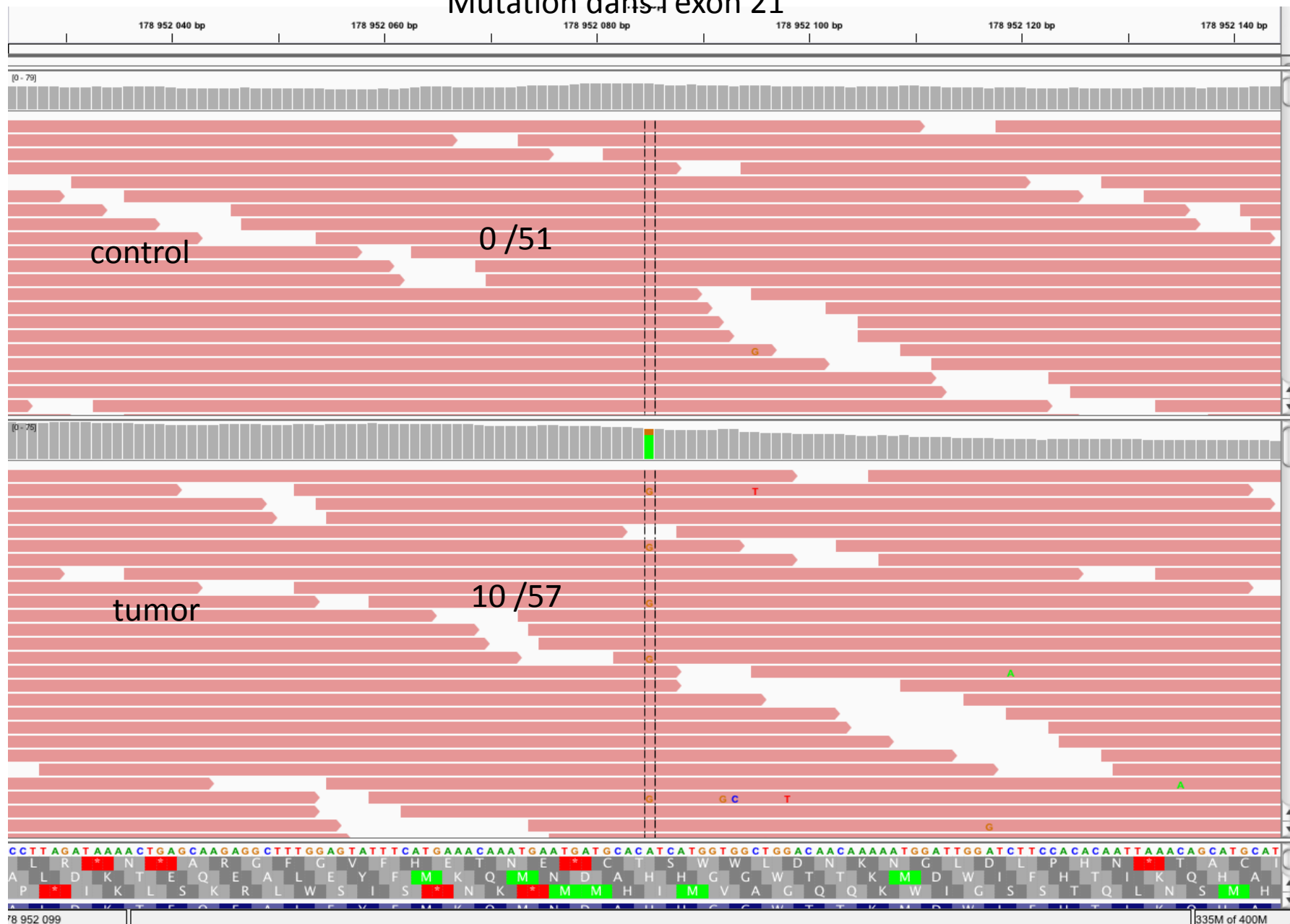


En 5' de l'exon 5

Ici reads simples orientés (par de pair-end :
1 seule couleur rouge)



Mutation dans l'exon 21



**Test statistique de Fisher
pour évaluer la pertinence de la mutation**

Utilisation de la fonction `fisher.test` de R

Test exact de Fisher

Le **Test exact de Fisher** est un test statistique utilisé pour l'analyse des tables de contingence. Ce test est utilisé en général avec des faibles effectifs mais il est valide pour toutes les tailles d'échantillon. Il doit son nom à son inventeur [Ronald Fisher](#). C'est un test qualifié d'exact car les probabilités peuvent être calculées exactement plutôt qu'en s'appuyant sur une approximation qui ne devient correcte qu'asymptotiquement comme pour le test du χ^2 utilisé dans les tables de contingence.

Les calculs à la main ne sont raisonnables que pour les tables 2 × 2 mais le principe du test peut s'étendre au cas général et certains logiciels de statistique permettent le calcul pour le cas général.

Exemple [\[modifier \]](#) [modifier le code](#)

Soit un échantillon d'adolescents on sépare l'échantillon entre filles et garçons et entre ceux qui suivent un régime et ceux qui n'en suivent pas et nous supposons que la proportion de filles qui suivent un régime est supérieure à celle des garçons, et nous voulons tester si la différence de proportions observées est significative. Voici les données

	Garçons	Filles	<i>total ligne</i>
régime	1	9	10
non régime	11	3	14
<i>total colonne</i>	12	12	24

Ces données ne sont pas adaptées pour une analyse par un test du χ^2 , parce que les valeurs attendues (théoriques) dans la table sont inférieures à 10, et dans une table de contingence 2 × 2, le nombre de degrés de liberté est toujours égal à 1.

La question que l'on se pose à propos de ces données est : sachant que 10 de ces 24 adolescents pratiquent un régime et que 12 sont des filles, quelle est la probabilité que ces 10 qui pratiquent un régime soient répartis de manière équilibrée entre les filles et les garçons ? Si on choisit 10 adolescents au hasard, quelle est la probabilité que 9 d'entre eux soient parmi les 12 filles et seulement 1 parmi les 12 garçons ?

Avant de passer au test de Fisher nous introduisons quelques notations. On représente les cellules par les lettres *a*, *b*, *c* et *d* et on note *n* le total général. La table se présente ainsi :

	Garçons	Filles	<i>Total</i>
Régime	<i>a</i>	<i>b</i>	<i>a + b</i>
Non-régime	<i>c</i>	<i>d</i>	<i>c + d</i>
<i>Totaux</i>	<i>a + c</i>	<i>b + d</i>	<i>a + b + c + d (=n)</i>

Fisher a montré que la probabilité d'obtenir un tel ensemble de valeurs était donnée par la loi hypergéométrique :

$$p = \frac{\binom{a+b}{a} \binom{c+d}{c}}{\binom{n}{a+c}} = \frac{(a+b)!(c+d)!(a+c)!(b+d)!}{a!b!c!d!n!}$$

où $\binom{n}{k}$ est le coefficient binomial et le symbole ! indique la factorielle.

Dans l'exemple la probabilité d'obtenir le tableau croisé observé, avec les totaux marginaux donnés, est donc:

$$p = \frac{10!14!12!12!}{1!9!11!3!24!} = \frac{40}{29716} = 0,001346...$$

De manière à calculer si des données observées sont significativement éloignées de l'indépendance, c'est-à-dire la probabilité d'observer des données aussi ou plus éloignées que celles observées si l'hypothèse nulle (indépendance) est satisfaite, il faut calculer les valeurs de p pour ces tables et les ajouter. Cela donne un test unilatéral; pour un test bilatéral on doit considérer les tables qui sont extrêmes mais dans l'autre direction. Malheureusement classer les tables pour savoir si elles sont ou non aussi extrêmes est problématique. Une approche utilisée dans R avec la fonction "fisher.test" calcule la valeur p en sommant les probabilités de toutes les tables ayant une probabilité inférieure ou égale à celle de la table observée.

Voici la commande permettant d'entrer le tableau croisé de l'exemple présenté plus haut comme une matrice de R et de calculer la valeur de la probabilité.

```
fisher.test(matrix(c(1,9,11,3),nrow=2))
```

Et voici la valeur de la probabilité obtenue:

```
p-value = 0.002759
```

Cette valeur se calcule aussi assez facilement sans logiciel puissant avec le principe exposé. Il y a en tout quatre tables aussi éloignées ou plus éloignées de l'indépendance que la table observée. Il n'y a en tout que onze tables possibles sachant qu'il y a seulement dix adolescents qui suivent un régime, le nombre de garçons suivant un régime peut varier de 0 jusqu'à 10. Une fois que le nombre de garçons suivant un régime est connu tout le reste de la table est connu, ce qui correspond intuitivement au degré de liberté valant 1. Les quatre tables extrêmes correspondent aux valeurs 0,1,9 et 10 pour le nombre de garçons suivant un régime. La probabilité des deux tables plus éloignées de l'indépendance est très faible elle se calcule comme précédemment:

$$p = \frac{10!14!12!12!}{0!10!12!2!24!} = \frac{1}{29716} = 0,0000336519...$$

En ajoutant les probabilités des quatre tables on trouve $p = \frac{82}{29716} = 0,0027594561...$

Le test permet de rejeter l'indépendance entre le sexe et le fait de faire un régime.

Fisher's Exact Test for Count Data

Usage

```
fisher.test(x, y = NULL, workspace = 200000, hybrid = FALSE,  
            control = list(), or = 1, alternative = "two.sided",  
            conf.int = TRUE, conf.level = 0.95,  
            simulate.p.value = FALSE, B = 2000)
```

Arguments

x either a two-dimensional contingency table in matrix form, or a factor object.

y a factor object; ignored if x is a matrix.

workspace an integer specifying the size of the workspace used in the network algorithm. In units of 4 bytes. Only used for non-simulated p-values larger than 2 by 2 tables.

hybrid a logical. Only used for larger than 2 by 2 tables, in which cases it indicated whether the exact probabilities (default) or a hybrid approximation thereof should be computed. See 'Details'.

control a list with named components for low level algorithm control. At present the only one used is "mult", a positive integer ≥ 2 with default 30 used only for larger than 2 by 2 tables. This says how many times as much space should be allocated to paths as to keys: see file 'fexact.c' in the sources of this package.

or the hypothesized odds ratio. Only used in the 2 by 2 case.

alternative indicates the alternative hypothesis and must be one of "two.sided", "greater" or "less". You can specify just the initial letter. Only used in the 2 by 2 case.

conf.int logical indicating if a confidence interval should be computed (and returned).

conf.level confidence level for the returned confidence interval. Only used in the 2 by 2 case if conf.int = TRUE.

simulate.p.value a logical indicating whether to compute p-values by Monte Carlo simulation, in larger than 2 by 2 tables

B an integer specifying the number of replicates used in the Monte Carlo test.

A list with class "htest" containing the following components:

p.value the p-value of the test.

conf.int a confidence interval for the odds ratio. Only present in the 2 by 2 case if argument conf.int = TRUE.

estimate an estimate of the odds ratio. Note that the *conditional* Maximum Likelihood Estimate (MLE) rather than the unconditional MLE (the sample odds ratio) is used. Only present in the 2 by 2 case.

null.value the odds ratio under the null, or. Only present in the 2 by 2 case.

alternative a character string describing the alternative hypothesis.

method the character string "Fisher's Exact Test for Count Data".

data.name a character string giving the names of the data.

	control	tumor	
A	a	b	a+b
G	c	d	c+d
Total	a+c	b+d	a+b+c+d
	control	tumor	
A	51	47	98
G	0	10	10
Total	51	57	108

```
> m <- matrix(c(51,47,0,10), byrow=T,
nrow = 2, dimnames = list(c("A","G"),
c("Control", "Tumor")))
```

```
> m
```

```
      Control Tumor
A      51     47
G       0     10
```

```
> fisher.test(m)$p.value
[1] 0.001445162
```

ou plus simplement (en une seule ligne) :

```
fisher.test(matrix(c(51,47,0,10),byrow=T,nr
ow = 2))$p.value
```

La probabilité de la mutation A>G est dans ces conditions de $1.4 \cdot 10^{-3}$

G n'est pas présent dans le control. Il apparaît à 10/57 dans la tumeur

La mutation n affecte pas 100% des reads .

Probablement mutation hétérozygote ... et % cellules tumorales < 100%

En général on filtrera les variants entre normal et tumoral à > 0.01 .

Digression sur le logiciel R

www.r-project.org

The R Project for Statistical Computing



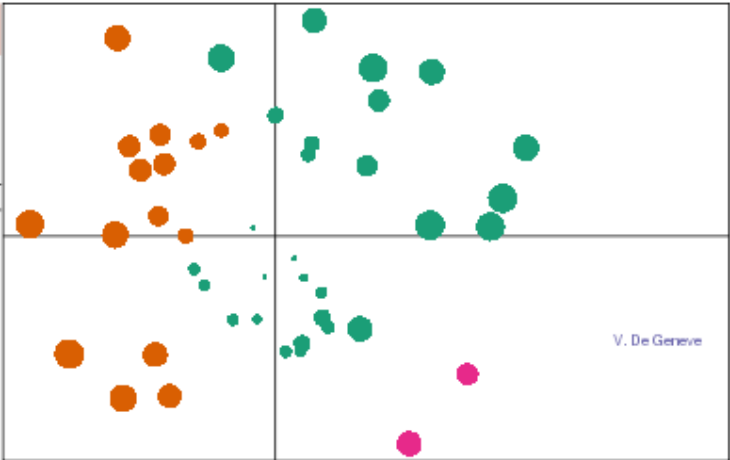
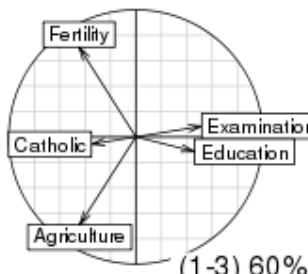
About R
[What is R?](#)
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[Screenshots](#)
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Download, Packages
[CRAN](#)

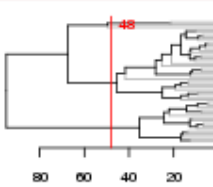
R Project
[Foundation](#)
[Members &](#)
[Donors](#)
[Mailing Lists](#)
[Bug Tracking](#)
[Developer Page](#)
[Conferences](#)
[Search](#)

Documentation
[Manuals](#)

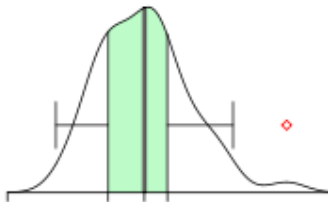
PCA 5 vars
`princomp(x = data, cor = cor)`



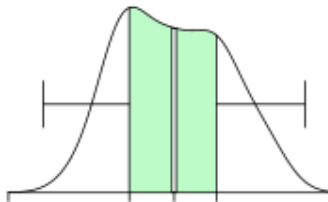
Clustering 4 groups



Factor 1 [41%]



Factor 3 [19%]



Getting Started:

- R is a free software environment for statistical computing and graphics. It compiles and runs on a wide variety of UNIX platforms, Windows and MacOS. To [download R](#), please choose your preferred [CRAN mirror](#).

Logiciel (sur PC, Mac Unix) d analyse statistique et graphique



The Comprehensive R Archive Network

Download and Install R

Precompiled binary distributions of the base system and contributed packages, **Windows and Mac** users most likely want one of these versions of R:

- [Download R for Linux](#)
- [Download R for \(Mac\) OS X](#)
- [Download R for Windows](#)

R is part of many Linux distributions, you should check with your Linux package management system in addition to the link above.

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Manuals



edited by the R Development Core Team.

The following manuals for R were created on Debian Linux and may differ from the manuals for Mac or Windows on platform-specific pages, but most parts will be identical for all platforms. The correct version of the manuals for each platform are part of the respective R installations. The manuals change with R, hence we provide versions for the most recent released R version (R-release), a very current version for the patched release version (R-patched) and finally a version for the forthcoming R version that is still in development (R-devel).

Here they can be downloaded as PDF files, EPUB files, or directly browsed as HTML:

[CRAN](#)
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Manual	R-release	R-patched	R-devel
An Introduction to R is based on the former "Notes on R", gives an introduction to the language and how to use R for doing statistical analysis and graphics.	HTML PDF EPUB	HTML PDF EPUB	HTML PDF EPUB
R Data Import/Export describes the import and export facilities available either in R itself or via packages which are available from CRAN.	HTML PDF EPUB	HTML PDF EPUB	HTML PDF EPUB
R Installation and Administration	HTML PDF EPUB	HTML PDF EPUB	HTML PDF EPUB

<https://pbil.univ-lyon1.fr/CRAN/>

2. An Overview of R.....

2.1 The Uses of R.....

2.1.1 R may be used as a calculator.....

2.1.2 R will provide numerical or graphical summaries of data

2.1.3 R has extensive graphical abilities

2.1.4 R will handle a variety of specific analyses

2.1.5 R is an Interactive Programming Language

“R for Beginners” by Emmanuel Paradis ([PDF](#)).

http://cran.univ-lyon1.fr/doc/contrib/Paradis-rdebuts_en.pdf

“Using R for Data Analysis and Graphics - Introduction, Examples and Commentary” by John Maindonald

<http://cran.univ-lyon1.fr/doc/contrib/usingR.pdf>

Type 'demo()' for some demos, 'help()' for on-line help, or
'help.start()' for an HTML browser interface to help.
Type 'q()' to quit R.

```
> 2+2
```

```
[1] 4
```

```
> x <- 10
```

```
> x
```

```
[1] 10
```

```
> x <- rep(1:10,3)
```

```
> mean(x)
```

```
[1] 5.5
```

```
> sd(x)
```

```
[1] 2.921384
```

```
> fisher.test(matrix(c(51,47,0,10), byrow=T, nrow = 2))$p.value
```

On

```
fisher.test(matrix(c(51,0,47,10), nrow = 2))$p.value
```

```
> help.start()
```

```
> help(plot)
```

R: Documentation x The R Language x +

127.0.0.1:19533/doc/html/index.html Ask Search

Statistical Data Analysis

Manuals

An Introduction to R	The R Language Definition
Writing R Extensions	R Installation and Administration
R Data Import/Export	R Internals

Reference

Packages	Search Engine & Keywords
--------------------------	--

Miscellaneous Material

About R	Authors	Resources
License	Frequently Asked Questions	Thanks
NEWS	User Manuals	Technical papers

Material specific to the Windows port

CHANGES up to R 2.15.0	Windows FAQ
--	-----------------------------

>help.start()

Caractérisation de la mutation H 1047 R dans la PIK3CA dans les bases de données

Cosmic » Gene » Overview » [PIK3CA](#)

[Overview](#)
[Sequence](#)
[Fusions](#)
[Studies](#)
[References](#)

Gene name [PIK3CA](#)

Genomic Coordinates [3:178916614..178952152](#)

Gene Id COSG14

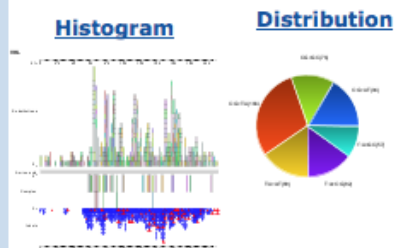
Synonyms MGC142161,MGC142163,PI3K,p110-alpha,P42336,ENSG00000121879

Drug Sensitivity Data: Mutations in PIK3CA are associated with altered sensitivity to the following drug(s) : [AZD6482](#), [Bexarotene](#), [AKT inhibitor VIII](#), [WQ2009093972](#), [AMG-706](#), [CI-1040](#), [PAC-1](#), [WZ-1-84](#), [Lapatinib](#), [AZD6482](#), [AKT inhibitor VIII](#), [AZD6482](#), [Methotrexate](#), [VX-702](#), [PAC-1](#)
 Click [here](#) for PIK3CA all drug sensitivity data

No.of Samples Total number of unique samples: 72354
 Unique samples with mutations: 7418

Alternative Transcripts [PIK3CA_ENST00000263967](#)

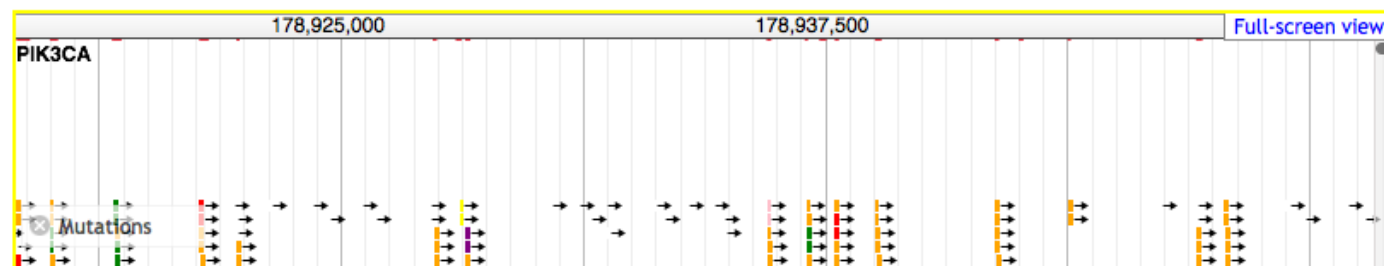
Mutation Analysis



External Links

OMIM: [171834](#)
Transcript: [NM_006218.1](#)
Ensembl Contig View: [PIK3CA](#)
UCSC Browser: [PIK3CA](#)
Copy Number: [CONAN](#)
Pfam: [P42336](#)
Atlas Genetic [GC_PIK3CA](#)
Oncology:
HGNC: [8975](#)

Cosmic Genome Browser



*Cosmic » Mutation » Overview » **PIK3CA** » **p.H1047R / c.3140A>G***

Overview Tissue Distribution Samples Pathways Affected References

Gene Name: PIK3CA

Mutation Id: COSM775

AA Mutation: p.H1047R (Substitution - Missense)

CDS Mutation: c.3140A>G (Substitution)

GRCh37: [Ensembl Contig View](#) 3:178952080..178952090

COSMIC Genome Browser: [COSMIC JBrowse](#) 3:178952080..178952090

CDD: [NP_006209.2](#)

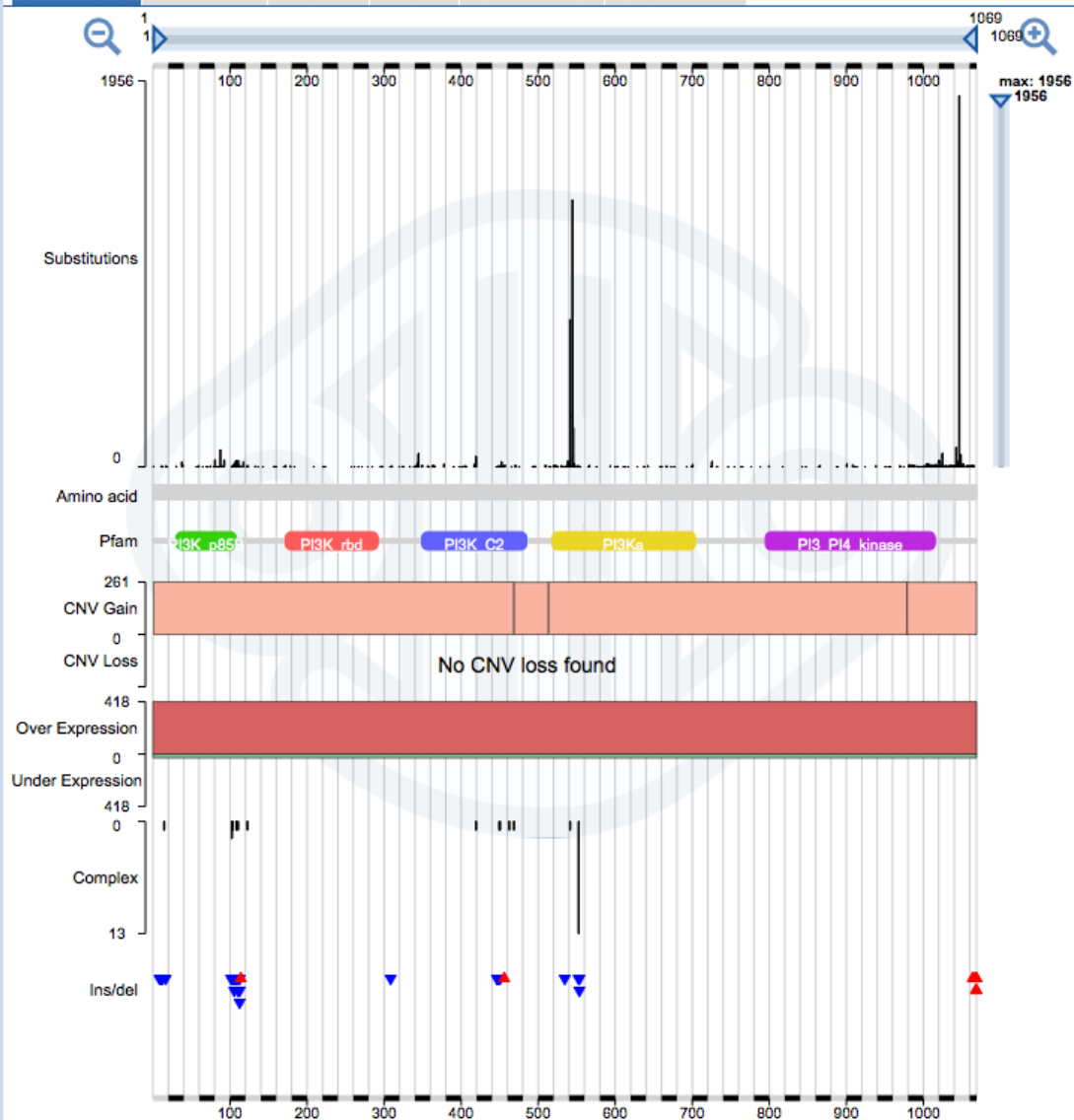
Homologene: [21249](#), click [here](#) to look at the multiple sequence alignment.

Ever confirmed somatic: Yes

PIK3CA(c.3045T>C;p.S1015S)
 ↳ PIK3CA_ENST00000263967(c.3045T>C;p.S1015S)
 ↳ PIK3CA(c.3046T>C;p.F1016L)
 ↳ PIK3CA_ENST00000263967(c.3046T>C;p.F1016L)
 ↳ PIK3CA(c.3047T>G;p.F1016C)
 ↳ PIK3CA_ENST00000263967(c.3047T>G;p.F1016C)
 ↳ PIK3CA(c.3049G>C;p.D1017H)
 ↳ PIK3CA_ENST00000263967(c.3049G>C;p.D1017H)
 ↳ PIK3CA(c.3050A>G;p.D1017G)
 ↳ PIK3CA(c.3050A>T;p.D1017V)
 ↳ PIK3CA_ENST00000263967(c.3050A>G;p.D1017G)
 ↳ PIK3CA(c.3051T>G;p.D1017E)
 ↳ PIK3CA_ENST00000263967(c.3051T>G;p.D1017E)
 ↳ PIK3CA(c.3051T>C;p.D1017D)
 ↳ PIK3CA_ENST00000263967(c.3051T>C;p.D1017D)
 ↳ PIK3CA(c.3052G>A;p.D1018N)
 ↳ PIK3CA_ENST00000263967(c.3052G>A;p.D1018N)
 ↳ PIK3CA(c.3053A>G;p.D1018G)
 ↳ PIK3CA_ENST00000263967(c.3053A>G;p.D1018G)
 ↳ PIK3CA(c.3054C>T;p.D1018D)
 ↳ PIK3CA_ENST00000263967(c.3054C>T;p.D1018D)
 ↳ PIK3CA(c.3142C>T;p.H1048Y)
 ↳ PIK3CA_ENST00000263967(c.3142C>A;p.H1048N)
 ↳ PIK3CA(c.3143A>G;p.H1048R)
 ↳ PIK3CA(c.3143A>T;p.H1048L)
 ↳ PIK3CA_ENST00000263967(c.3143A>G;p.H1048R)
 ↳ PIK3CA(c.3144T>G;p.H1048Q)
 ↳ PIK3CA(c.3145G>A;p.G1049S)
 ↳ PIK3CA(c.3145G>C;p.G1049R)
 ↳ PIK3CA(c.3145G>T;p.G1049C)
 ↳ PIK3CA_ENST00000263967(c.3145G>A;p.G1049S)
 ↳ PIK3CA_ENST00000263967(c.3145G>C;p.G1049R)
 ↳ PIK3CA(c.3146G>A;p.G1049D)
 ↳ PIK3CA(c.3146G>C;p.G1049A)
 ↳ PIK3CA_ENST00000263967(c.3146G>A;p.G1049D)
 ↳ PIK3CA_ENST00000263967(c.3146G>C;p.G1049A)

Cosmic » Gene » Analysis » PIK3CA

Histogram Mutations Fusions Tissue Distribution CNV & Expr



Filters

Gene

Position

Start

End

Sequence Type:

cDNA ☐

Amino Acid ☒

☒ Systematic Screen

☒ Somatic Status

☒ Tumour Source

☒ Mutation Type

☒ Copy Number Variation

☒ Gene Expression

Apply

Reset

Cosmic » *Gene* » *Analysis* » ***PIK3CA***

[Histogram](#)
[Mutations](#)
[Fusions](#)
[Tissue](#)
[Distribution](#)
[CNV & Expr](#)

Show entries

Search:

Export: [CSV](#) [TSV](#)

Position (AA) ^	Mutation (CDS) v	Mutation (Amino Acid) v	Mutation ID (COSM) v	Count v	Mutation Type v
1	c.1A>G	p.M1V	COSM1041443	4	Substitution - Missense
9	c.24_56del33	p.E9_R19del	COSM39613	1	Deletion - In frame
10	c.28_48del21	p.L10_M16del	COSM35324	1	Deletion - In frame
11	c.30_53del24	p.W11_P18delWGIHLMPP	COSM1041445	1	Deletion - In frame
11	c.32G>T	p.W11L	COSM582520	1	Substitution - Missense
12	c.35G>A	p.G12D	COSM27495	5	Substitution - Missense
13	c.36_56del21	p.I13_R19del	COSM166338	1	Deletion - In frame
13	c.38T>C	p.I13T	COSM1420724	1	Substitution - Missense
15	c.42_64>TCCAA	p.L15_V22>PM	COSM87211	1	Complex - deletion inframe
17	c.49C>T	p.P17S	COSM27494	1	Substitution - Missense

Showing 1 to 10 of 703 entries

[First](#)
[Previous](#)
[1](#)
[2](#)
[3](#)
[4](#)
[5](#)
[Next](#)
[Last](#)

Filters

Gene

Position

Start

End

Sequence Type:

[cDNA](#) ☐

[Amino Acid](#) ☒

☒ **Systematic Screen**

☒ **Somatic Status**

☒ **Tumour Source**

☒ **Mutation Type**

☒ **Copy Number Variation**

☒ **Gene Expression**

Apply

Reset

UNIPROT : acc P42336

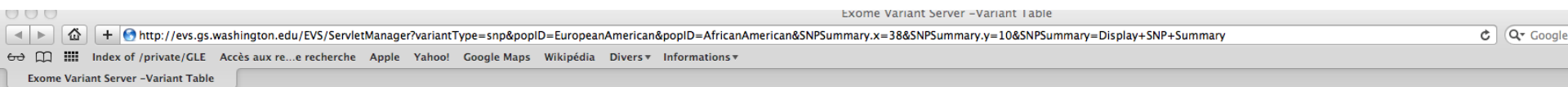
		activity. 2 Publications	
Natural variant ⁱ	1047 – 1047	1 H → R in CLOVE, KERSEB, CRC, BC and OC; also found in an endometrial carcinoma sample; shows an increase in lipid kinase activity; oncogenic in vivo; requires binding to p85 regulatory subunit to induce cellular transformation but not interaction with RAS; may mimic the conformatitonal change triggered by the interaction with RAS; enhances invadopodia-mediated extracellular matrix degradation and invasion in breast cancer cells; increases lipid kinase activity; may alter the interaction of the PI3K/PI4K kinase domain with the cell membrane. 12 Publications	VAR_026192

Breast cancer (BC) [MIM:114480]: A common malignancy originating from breast epithelial tissue. Breast neoplasms can be distinguished by their histologic pattern. Invasive ductal carcinoma is by far the most common type. Breast cancer is etiologically and genetically heterogeneous. Important genetic factors have been indicated by familial occurrence and bilateral involvement. Mutations at more than one locus can be involved in different families or even in the same case. 1 Publication

Note: Disease susceptibility is associated with variations affecting the gene represented in this entry.

Exome variant Server

http://evs.gs.washington.edu/EVS/



Gene Name: [PIK3CA](#) (Gene ID: 5290) (+)

Chromosome 3: 178866311 - 178952500

Population: EuropeanAmerican, AfricanAmerican

GWAS Catalog: [PIK3CA](#)

KEGG Pathway: [PIK3CA](#)

Sanger COSMIC: [PIK3CA](#)

PPI STRING 9.0: [PIK3CA](#)

OMIM: [PIK3CA](#)

Variation Color Code:

- splice or nonsense or frameshift
- missense
- coding-synonymous
- coding
- utr
- codingComplex

Download Option:

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Zip Format [gzip](#)

[download](#)

Add or Remove Columns ([Description of Columns](#))

☒ dbSNP rs ID	☒ Alleles	☒ EA Allele Count	☒ AA Allele Count	☒ Allele Count	☒ EA Genotype Count	☒ AA Genotype Count
☒ Genotype Count	☐ MAF (%)	☒ Sample Read Depth	☒ Genes	☒ Gene Accession #	☒ GVS Function	☒ cDNA Change
☒ cDNA Size	☒ Protein Change	☒ Conservation (GERP)	☐ Conservation (phastCons)	☒ Grantham Score	☒ PolyPhen Prediction	☐ Clinical Link
☐ NCBI 37 Allele	☐ Chimp Allele	☐ Illumina HumanExome Chip	☐ GWAS Hits	☐ EA Est. Age (kyrs)	☐ AA Est. Age (kyrs)	☐ GRCh38 Position

Sort Variants by [Variant Pos](#)

Select Population [All](#)

Select Transcript [NM_006218.2](#)

If "Select Transcript" above is set to "Union of Transcripts", and if multiple transcripts of a gene are involved in a variant and the function annotations for the variant are the same, only one representative transcript is listed in the table below for the reasons of speed and space. But annotations for each individual transcript are available in the full report. [download the data.](#)

Show 10 entries																
Variant GRCh37 Pos	rs ID	Alleles	EA Allele #	AA Allele #	All Allele #	EA Genotype #	AA Genotype #	All Genotype #	Avg. Sample Read Depth	Genes	mRNA Accession #	GVS Function	cDNA Change	cDNA Size	Protein Change	C
3:178947078	rs202175608	T>C	C=3/T=8247	C=0/T=3790	C=3/T=12037	CC=0/CT=3/TT=4122	CC=0/CT=0/TT=1895	CC=0/CT=3/TT=6017	143	PIK3CA	NM_006218.2	coding-synonymous	c.2514T>C	3207	p.(C838=)	2
3:178947241	rs201113847	T>G	G=4/T=8216	G=0/T=3764	G=4/T=11980	GG=0/GT=4/TT=4106	GG=0/GT=0/TT=1882	GG=0/GT=4/TT=5988	66	PIK3CA	NM_006218.2	intron	c.2666+11T>G	3207	NA	-
3:178947256	rs371541151	A>C	C=0/A=8202	C=2/A=3726	C=2/A=11928	CC=0/CA=0/AA=4101	CC=0/CA=2/AA=1862	CC=0/CA=2/AA=5963	47	PIK3CA	NM_006218.2	intron	c.2666+26A>C	3207	NA	-
3:178947767	rs375882334	C>A	A=0/C=8198	A=1/C=3731	A=1/C=11929	AA=0/AC=1/CC=4099	AA=0/AC=1/CC=1865	AA=0/AC=1/CC=5964	170	PIK3CA	NM_006218.2	intron	c.2667-25C>A	3207	NA	3
3:178948147	rs369770518	G>A	A=1/G=8155	A=0/G=3618	A=1/G=11773	AA=0/AG=1/GG=4077	AA=0/AG=0/GG=1809	AA=0/AG=1/GG=5886	70	PIK3CA	NM_006218.2	coding-synonymous	c.2919G>A	3207	p.(K973=)	-
3:178948172	rs372832719	C>T	T=1/C=8139	T=0/C=3610	T=1/C=11749	TT=0/TC=1/CC=4069	TT=0/TC=0/CC=1805	TT=0/TC=1/CC=5874	59	PIK3CA	NM_006218.2	intron	c.2936+8C>T	3207	NA	-
3:178948196	rs3729693	A>G	G=1/A=8139	G=452/A=3168	G=453/A=11307	GG=0/GA=1/AA=4069	GG=27/GA=398/AA=1385	GG=27/GA=399/AA=5454	39	PIK3CA	NM_006218.2	intron	c.2936+32A>G	3207	NA	2
3:178951894	rs369697984	G>A	A=0/G=8190	A=1/G=3729	A=1/G=11919	AA=0/AG=0/GG=4095	AA=0/AG=1/GG=1864	AA=0/AG=1/GG=5959	111	PIK3CA	NM_006218.2	missense	c.2949G>A	3207	p.(M983I)	6
3:178951926	rs373295359	A>G	G=1/A=8211	G=0/A=3750	G=1/A=11961	GG=0/GA=1/AA=4105	GG=0/GA=0/AA=1875	GG=0/GA=1/AA=5980	116	PIK3CA	NM_006218.2	missense	c.2981A>G	3207	p.(H994R)	6
3:178952020	rs17849079	C>T	T=36/C=8204	T=104/C=3648	T=140/C=11852	TT=0/TC=36/CC=4084	TT=2/TC=100/CC=1774	TT=2/TC=136/CC=5858	90	PIK3CA	NM_006218.2	coding-synonymous	c.3075C>T	3207	p.(T1025=)	2

Showing 81 to 90 of 91 entries

*In general, the INDEL calls are less robust than the SNP calls and have a higher false positive rate. When applying the ESP data to research studies, users are advised to keep this difference in mind.

~ sign in front of a dbSNP rsID indicates an INDEL is approximately mapped to the rsID by [SeattleSeqAnnotation137](#). It should be considered as a suggestion rather than an accurate mapping to the existing records in dbSNP.



Visualize, analyze, discover.



Memorial Sloan Kettering Cancer Center.

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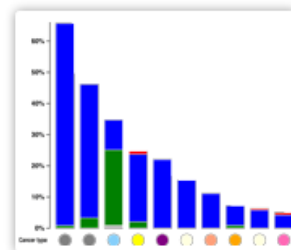
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VISUALIZE YOUR DATA

The cBioPortal for Cancer Genomics provides **visualization, analysis** and **download** of large-scale **cancer genomics** data sets.

Please adhere to [the TCGA publication guidelines](#) when using TCGA data in your publications.

Please cite Gao et al. *Sci. Signal.* 2013 & Cerami et al. *Cancer Discov.* 2012 when publishing results based on cBioPortal.



Query

Download Data

Select Cancer Study:

All Cancer Studies

Select Data Type Priority:

☒ Mutation and CNA ☐ Only Mutation ☐ Only CNA

Enter Gene Set:

Advanced: Onco Query Language (OQL)

User-defined List

PIK3CA

☒ All gene symbols are valid.

Submit

http://www.cbioportal.org/index.do

What's New

- We are hiring a **Front End Engineer**
- We are hiring a **Data Curator**
- **New data and features released**

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Data Sets

The Portal contains data for **17584 tumor samples from 69 cancer studies**. [\[Details.\]](#)



Example Queries

RAS/RAF alterations in colorectal cancer

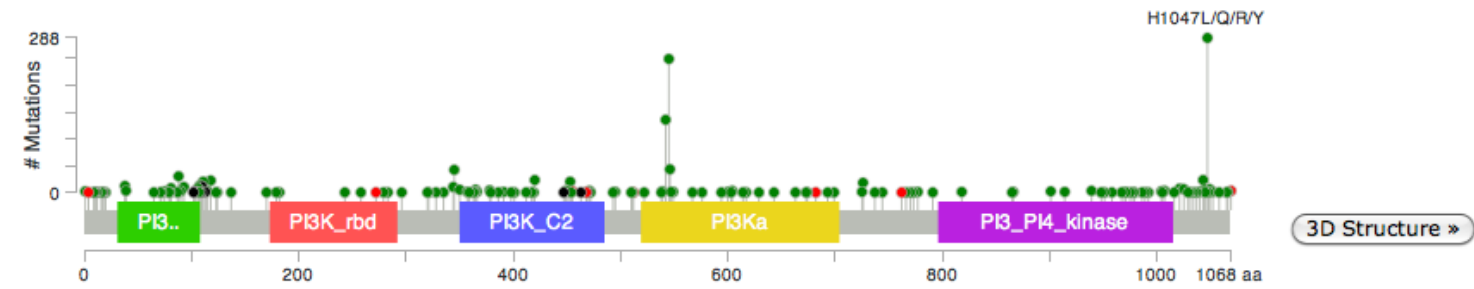
BRCA1 and BRCA2 mutations in ovarian cancer

POLE hotspot mutations in endometrial cancer

Multiple alterations

PIK3CA:

PK3CA_HUMAN



Showing 1,750 mutation(s)										
Search:										
Case ID	Cancer Study	AA change	Type	Copy #	COSMIC	VS	Mutation Assessor	Allele Freq (T)	#Mut in Sample	
SA217	Breast (BCCRC)	103_106EPVG>D 3D	IF del	NA	2	U		NA	16	
TCGA-A2-A0EV	Breast (TCGA pub)	469_469E>DK 3D	IF ins	gain	1	U		NA	46	
TCGA-B6-A0WY	Breast (TCGA pub)	109_112EEKI>D 3D	IF del	gain	1	U		NA	30	
TCGA-A2-A0EV	Breast (TCGA)	469_469E>DK 3D	IF ins	gain	1	U		NA	46	
KYSE70_OESOPHAGUS	CCLE	*1069W	Nonstop	AMP	1	U		0.86	63	
IGROV1_OVARY	CCLE	*1069W	Nonstop	diploid	1	U		0.52	276	
TCGA-32-1979	GBM (TCGA 2013)	*1069W	Nonstop	diploid	1	U		0.35	53	
TCGA-06-0879	GBM (TCGA 2013)	455_460LLNPIG>F 3D	IF del	diploid		U		0.15	39	
TCGA-02-0046	GBM (TCGA 2008)	17_18PP>P 3D	IF del	diploid		V		NA	4	
TCGA-32-1979	GBM (TCGA)	*1069W	Nonstop	diploid	1	U		NA	71	
TCGA-06-0879	GBM (TCGA)	455_460LLNPIG>F 3D	IF del	diploid		U		NA	49	
TCGA-73-7499	Lung adeno (TCGA pub)	123_124MP>IA 3D	Missense	gain	1	U		0.36	77	
IGROV1	NCI-60	*1069W	Nonstop	diploid	1	U		NA	620	
PT143MRC	Sarcoma (MSKCC)	MUTATED	Missense	diploid		V		NA	1	
PT195SYN	Sarcoma (MSKCC)	MUTATED	FS del	diploid		V		NA	2	
PT173PL	Sarcoma (MSKCC)	MUTATED	FS del	gain		V		NA	2	
PT149MRC	Sarcoma (MSKCC)	MUTATED	Missense	diploid		V		NA	2	
PT138MRC	Sarcoma (MSKCC)	MUTATED	Missense	diploid		V		NA	1	
PT158MRC	Sarcoma (MSKCC)	MUTATED	Missense	diploid		V		NA	1	
TCGA-BG-A0MT	Uterine (TCGA)	463_465GSN>D 3D	IF del	gain	1	U		NA	41	
TCGA-AP-A051	Uterine (TCGA)	e1-1 3D	Splice	diploid		U		0.50	6542	
TCGA-BG-A0MT	Uterine (TCGA pub)	463_465GSN>D 3D	IF del	gain	1	U		NA	43	

PIK3CA:

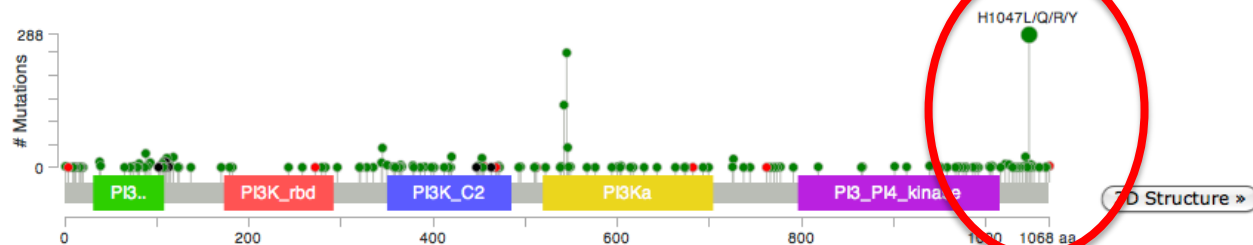
PK3CA_HUMAN

PDF

SVG

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Color Codes



Current view shows filtered results. Click [here](#) to reset all filters.

Showing 400 mutation(s) (out of 1,750 total mutations)										
Search:										
Case ID	Cancer Study	AA change	Type	Copy #	COSMIC	VS	Mutation Assessor	Allele Freq (T)	#Mut in Sample	
TCGA-OR-A5LK	ACC (TCGA)	H1047R	3D	Missense	gain	1878	U	Neutral	NA	82
BL018	Bladder (MSKCC 2012)	H1047R	3D	Missense	diploid	1878	V	Neutral	NA	1
BL067	Bladder (MSKCC 2012)	H1047R	3D	Missense	diploid	1878	V	Neutral	NA	1
B78	Bladder (BGI)	H1047R	3D	Missense	NA	1878	U	Neutral	NA	100
TCGA-FD-A3SJ	Bladder (TCGA pub)	H1047L	3D	Missense	gain	1878	U	Neutral	0.43	202
TCGA-HT-7478	Glioma (TCGA)	H1047L	3D	Missense	diploid	1878	U	Neutral	0.13	27
TCGA-FG-8187	Glioma (TCGA)	H1047R	3D	Missense	diploid	1878	U	Neutral	0.08	14
SA223	Breast (BCCRC)	H1047R	3D	Missense	NA	1878	U	Neutral	NA	15
SA218	Breast (BCCRC)	H1047R	3D	Missense	NA	1878	U	Neutral	NA	126
SA214	Breast (BCCRC)	H1047R	3D	Missense	NA	1878	U	Neutral	NA	193
SA215	Breast (BCCRC)	H1047R	3D	Missense	NA	1878	U	Neutral	NA	12
SA056	Breast (BCCRC)	H1047R	3D	Missense	NA	1878	U	Neutral	NA	14
BR-V-003	Breast (Broad)	H1047R	3D	Missense	NA	1878	V	Neutral	NA	13
BR-V-020	Breast (Broad)	H1047R	3D	Missense	NA	1878	U	Neutral	NA	15
BR-V-024	Breast (Broad)	H1047L	3D	Missense	NA	1878	V	Neutral	NA	26
BR-M-083	Breast (Broad)	H1047R	3D	Missense	NA	1878	U	Neutral	NA	17
BR-M-050	Breast (Broad)	H1047R	3D	Missense	NA	1878	V	Neutral	NA	24
BR-M-036	Breast (Broad)	H1047R	3D	Missense	NA	1878	V	Neutral	NA	11
BR-M-026	Breast (Broad)	H1047R	3D	Missense	NA	1878	V	Neutral	NA	27
BR-M-116	Breast (Broad)	H1047R	3D	Missense	NA	1878	V	Neutral	NA	102
BR-V-064	Breast (Broad)	H1047L	3D	Missense	NA	1878	U	Neutral	NA	17
BR-M-158	Breast (Broad)	H1047R	3D	Missense	NA	1878	V	Neutral	NA	16
BR-M-121	Breast (Broad)	H1047R	3D	Missense	NA	1878	U	Neutral	NA	26
BR-M-098	Breast (Broad)	H1047L	3D	Missense	NA	1878	V	Neutral	NA	29
BR-M-184	Breast (Broad)	H1047R	3D	Missense	NA	1878	V	Neutral	NA	11
BR-V-052	Breast (Broad)	H1047R	3D	Missense	NA	1878	V	Neutral	NA	28
PD4124a	Breast (Sanger)	H1047L	3D	Missense	NA	1878	U	Neutral	NA	28
PD4601a	Breast (Sanger)	H1047R	3D	Missense	NA	1878	U	Neutral	NA	151
PD4076a	Breast (Sanger)	H1047R	3D	Missense	NA	1878	U	Neutral	NA	20

BLAST mapping of all PDB structures to region PK3CA_HUMAN/797-1068

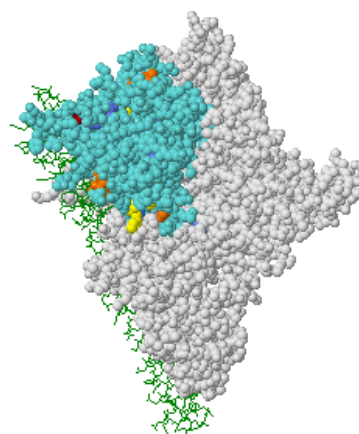
Only mappings with position of current variant **H1047R** shown

	PDB	e-value	length	identity	resolution	ligand	small.mol name	ligand protein ID	title
1	4a55:A	0	265	96.2%	3.5				crystal structure of p110alpha in complex with ish2 of p85alp the inhibitor pik-108
2	2rd0:A	0	254	96.1%	3.05				structure of a human p110alpha/p85alpha complex
3	3hbm:A	0	266	95.1%	2.8				crystal structure of p110alpha h1047r mutant in complex with nish2 of p85alpha and the drug wortmannin
4	3hiz:A	0	266	88.3%	3.3				crystal structure of p110alpha h1047r mutant in complex with nish2 of p85alpha

Mapped PDB chain shown with spacefill, all other chains - with wireframe. Structure centered on variant residue.

```
PK3CA_HUMAN/797-1061 : VMLGSGMPELQSFDIAYIRKTLALDKTEQEAL EYFMKQMND AHHGGWTTKMDWIFHT
midline : VMLGSGMPELQSFDIAYIRKTLALDKTEQEAL EYF KQMND AHHGGWTTKMDWIFHT
4a55:A/745-1001 : VMLGSGMPELQSFDIAYIRKTLALDKTEQEAL EYFTKQMND AHHGGWTTKMDWIFHT
```

specificity ■
conserved ■
neutral ■
unmapped ■
hetero ■
variant ■



1000 Genomes

http://phase3browser.1000genomes.org/Homo_sapiens/Gene/Variation_Gene/Table?db=core;g=ENSG00000121879;r=3:178865902-178957881#missense_variant_tablePanel

rs121913279	3:178952085	A/G/T	-	SNP	dbSNP			Missense variant	H/L	1047	0.75	0.085	ENST00000263967
rs121913279	3:178952085	A/G/T	-	SNP	dbSNP			Missense variant	H/R	1047	0.36	0.529	ENST00000263967
COSM776	3:178952085	A/T	-	somatic_SNV	COSMIC	-	-	Missense variant	H/L	1047	0.75	0.085	ENST00000263967
COSM775	3:178952085	A/G	-	somatic_SNV	COSMIC	-	-	Missense variant	H/R	1047	0.36	0.529	ENST00000263967
COSM249874	3:178952085	A/C	-	somatic_SNV	COSMIC	-	-	Missense variant	H/P	1047	0.21	0.825	ENST00000263967
COSM94987	3:178952085	A/T	-	somatic_SNV	COSMIC	-	-	Missense variant	H/L	1047	0.75	0.085	ENST00000263967
COSM94986	3:178952085	A/G	-	somatic_SNV	COSMIC	-	-	Missense variant	H/R	1047	0.36	0.529	ENST00000263967
COSM776	3:178952085	A/T	-	somatic_SNV	COSMIC	-	-	Missense variant	H/L	1047	0.75	0.085	ENST00000263967
COSM775	3:178952085	A/G	-	somatic_SNV	COSMIC	-	-	Missense variant	H/R	1047	0.36	0.529	ENST00000263967
COSM249874	3:178952085	A/C	-	somatic_SNV	COSMIC	-	-	Missense variant	H/P	1047	0.21	0.825	ENST00000263967
COSM94987	3:178952085	A/T	-	somatic_SNV	COSMIC	-	-	Missense variant	H/L	1047	0.75	0.085	ENST00000263967
COSM94986	3:178952085	A/G	-	somatic_SNV	COSMIC	-	-	Missense variant	H/R	1047	0.36	0.529	ENST00000263967

2-178,957,881

Gene: PIK3CA

Somatic mutation: COSM94986

COSM94986 SOMATIC SNV

Original source

Somatic mutations found in human cancers from the COSMIC project (release 67) | [View in COSMIC](#)

Alleles

A/G | Ancestral: **A** | Ambiguity code: **R**

Location

Chromosome **3:178952085** (forward strand) | [View in location tab](#)

Co-located

with **COSMIC** [COSM776](#) (A/T), [COSM775](#) (A/G), [COSM249874](#) (A/C), [COSM94987](#) (A/T) ; **dbSNP** [rs121913279](#) (A/G/T)

HGVS names ☐

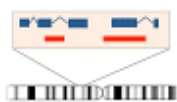
This variation has **3** HGVS names - click the plus to show

[3:g.178952085A>G](#)

[ENST00000263967.3:c.3140A>G](#)

[ENSP00000263967.3:p.His1047Arg](#)

Explore this variation



Genomic
context



Genes and
regulation



Sample
information



Individual
genotypes



Linkage
disequilibrium



Phenotype
data



Citations



Phylogenetic
context



Flanking
sequence

Original source	Somatic mutations found in human cancers from the COSMIC project (release 67) View in COSMIC
Alleles	A/G Ancestral: A Ambiguity code: R
Location	Chromosome 3:178952085 (forward strand) View in location tab
Co-located	with COSMIC COSM776 (A/T), COSM775 (A/G), COSM249874 (A/C), COSM94987 (A/T) ; dbSNP rs121913279 (A/G/T)
HGVS names 	This variation has 3 HGVS names - click the plus to show 3:g.178952085A>G ENST00000263967.3:c.3140A>G ENSP00000263967.3:p.His1047Arg

Phenotype Data

Significant association(s)

Show All  entries Show/hide columns Filter				
Disease/Trait	 Source(s)	 Tumour site	 Reported gene(s)	 Associated variant(s)
COSMIC tumour site: liver View on Karyotype	[COSMIC]	liver	PIK3CA ENST00000263967	COSM94986
COSMIC tumour site: skin View on Karyotype	[COSMIC]	skin	PIK3CA ENST00000263967	COSM94986
COSMIC tumour site: endometrium View on Karyotype	[COSMIC]	endometrium	PIK3CA ENST00000263967	COSM94986
COSMIC tumour site: breast View on Karyotype	[COSMIC]	breast	PIK3CA ENST00000263967	COSM94986
COSMIC tumour site: oesophagus View on Karyotype	[COSMIC]	oesophagus	PIK3CA ENST00000263967	COSM94986
COSMIC tumour site: stomach View on Karyotype	[COSMIC]	stomach	PIK3CA ENST00000263967	COSM94986
COSMIC tumour site: kidney View on Karyotype	[COSMIC]	kidney	PIK3CA ENST00000263967	COSM94986
COSMIC tumour site: lung View on Karyotype	[COSMIC]	lung	PIK3CA ENST00000263967	COSM94986
COSMIC tumour site: pancreas View on Karyotype	[COSMIC]	pancreas	PIK3CA ENST00000263967	COSM94986
COSMIC tumour site: central nervous system View on Karyotype	[COSMIC]	central nervous system	PIK3CA ENST00000263967	COSM94986
COSMIC tumour site: prostate View on Karyotype	[COSMIC]	prostate	PIK3CA ENST00000263967	COSM94986
COSMIC tumour site: ovary View on Karyotype	[COSMIC]	ovary	PIK3CA ENST00000263967	COSM94986
COSMIC tumour site: large intestine View on Karyotype	[COSMIC]	large intestine	PIK3CA ENST00000263967	COSM94986
COSMIC tumour site: urinary tract View on Karyotype	[COSMIC]	urinary tract	PIK3CA ENST00000263967	COSM94986
COSMIC tumour site: upper aerodigestive tract View on Karyotype	[COSMIC]	upper aerodigestive tract	PIK3CA ENST00000263967	COSM94986

PDBsum entry 2rd0



Go to PDB code:

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[Protein](#)

[Prot-prot](#)

[Pores](#)

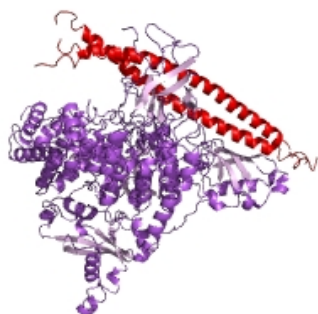
[Tunnels](#)

[Links](#)

Transferase/oncoprotein

PDB Id

2rd0



Contents

Protein chains

A 997 a.a. *

B 139 a.a. *

* Residue conservation analysis

PDB id: **2rd0**

[Links](#)

Name: **Transferase/oncoprotein**

Title: Structure of a human p110alpha/p85alpha complex

Structure: Phosphatidylinositol 4,5-bisphosphate 3-kinase ca subunit alpha isoform. Chain: a. Synonym: pi3-kinase p110 subunit alpha, ptdins-3- kinase p1 engineered: yes. Phosphatidylinositol 3-kinase regulatory subunit chain: b. Fragment: unp residues 322-600. Synonym: pi3-kinase p85 subunit alpha, ptdins-3-kinase p85-

Source: Homo sapiens. Human. Organism_taxid: 9606. Gene: pik3ca. Expressed in: spodoptera frugiperda. Expression_system_taxid: 7108. Gene: pik3r1, grb1.

Resolution: 3.05Å R-factor: 0.267 R-free: 0.323

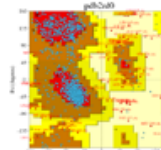
Authors: C.Huang,S.B.Gabelli,L.M.Amzel

Key ref: C.H.Huang et al. (2007). The structure of a human p110alpha/p85alpha complex elucidates the effects of oncogenic PI3Kalpha mutations. *Science*, 318, 1744-1748.

PubMed id: [18079394](#) DOI: [10.1126/science.1150799](#)

Date: 20-Sep-07 Release date: 25-Dec-07

PROCHECK



Headers

References



Protein chain **A**

P42336 (PK3CA_HUMAN) - Phosphatidylinositol 4,5-bisphosphate 3-kinase catalytic subunit alpha isoform

Seq:
Struc:

Seq:
Struc:

Seq: 1068 a.a.
Struc: 997 a.a. *

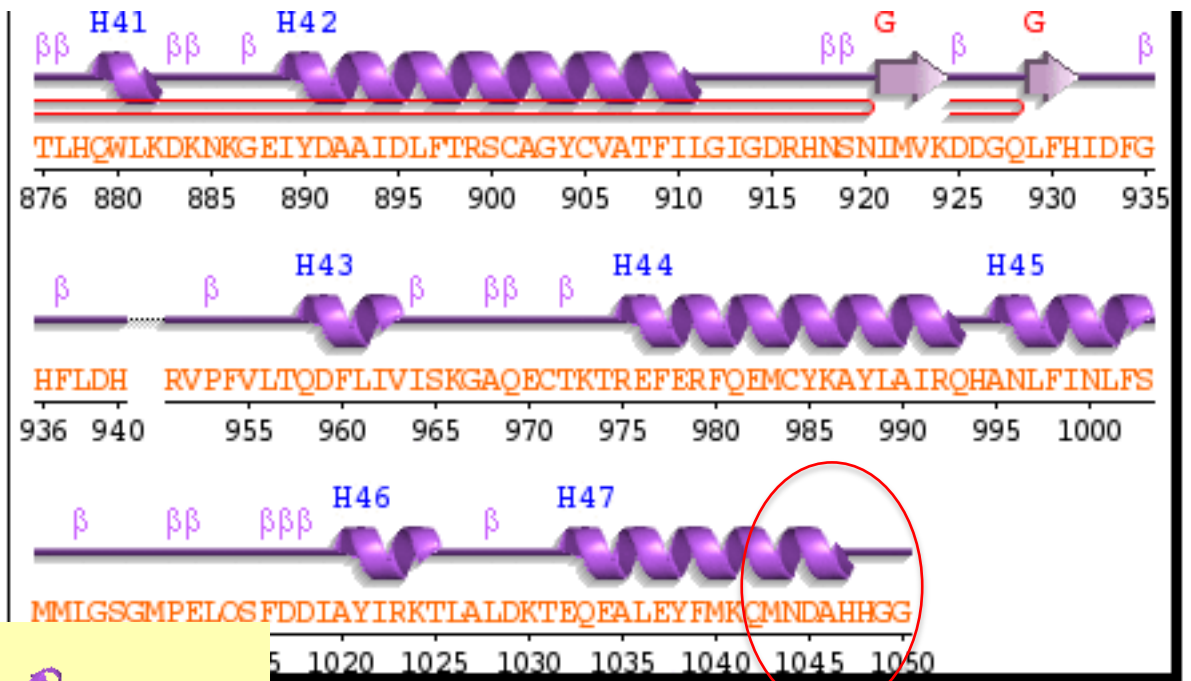
Protein chain **B**

P27986 (P85A_HUMAN) - Phosphatidylinositol 3-kinase regulatory subunit alpha

Seq:
Struc:

Seq: 724 a.a.
Struc: 139 a.a. *

Key: Family PfamA domain Secondary structure CATH domain
* PDB and UniProt seqs differ at 7 residue positions (black crosses)



 Helices labelled H1, H2, ... and strands by their sheets A, B, ...
 strand

